

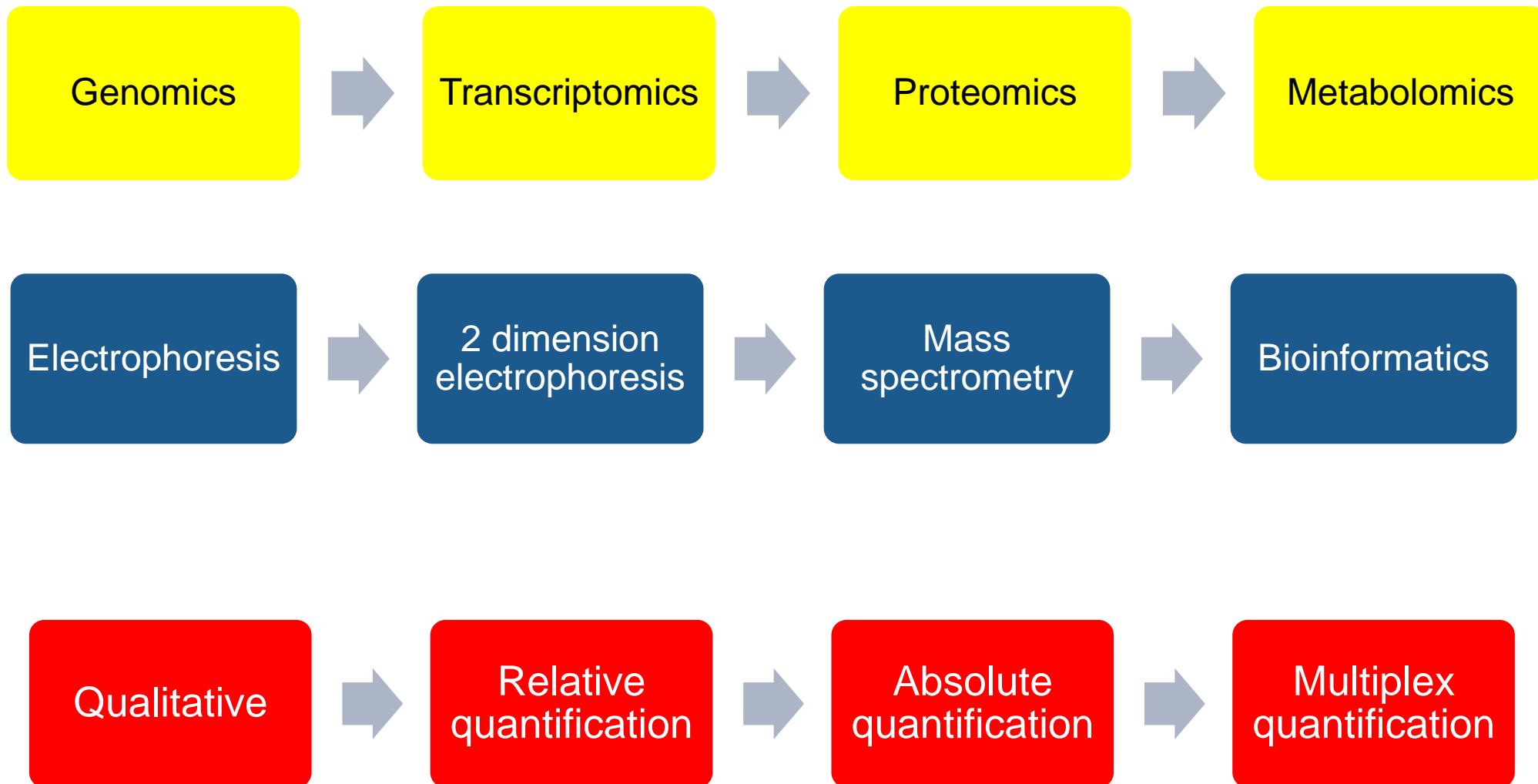
What Data Can We Get? The Potential for Omics in DairyCare

Professor David Eckersall

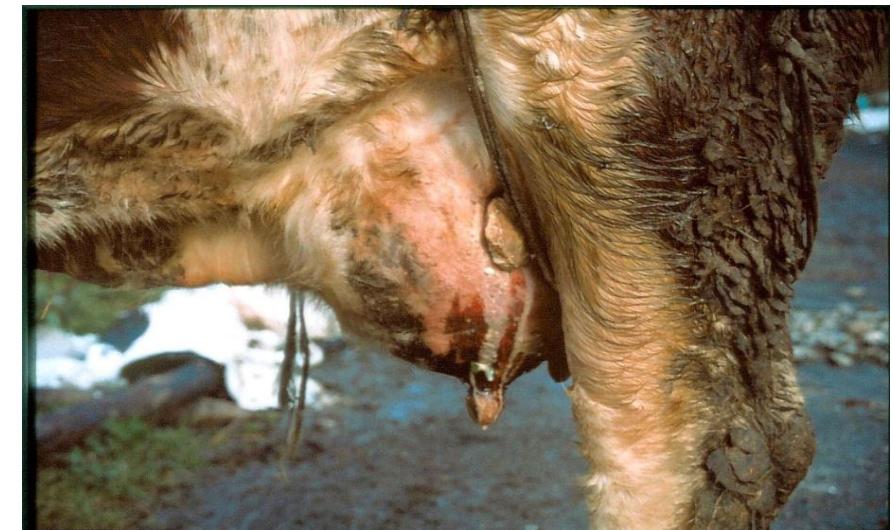
Veterinary Gene & Protein Group

**Institute of Biodiversity, Animal Health & Comparative
Medicine**

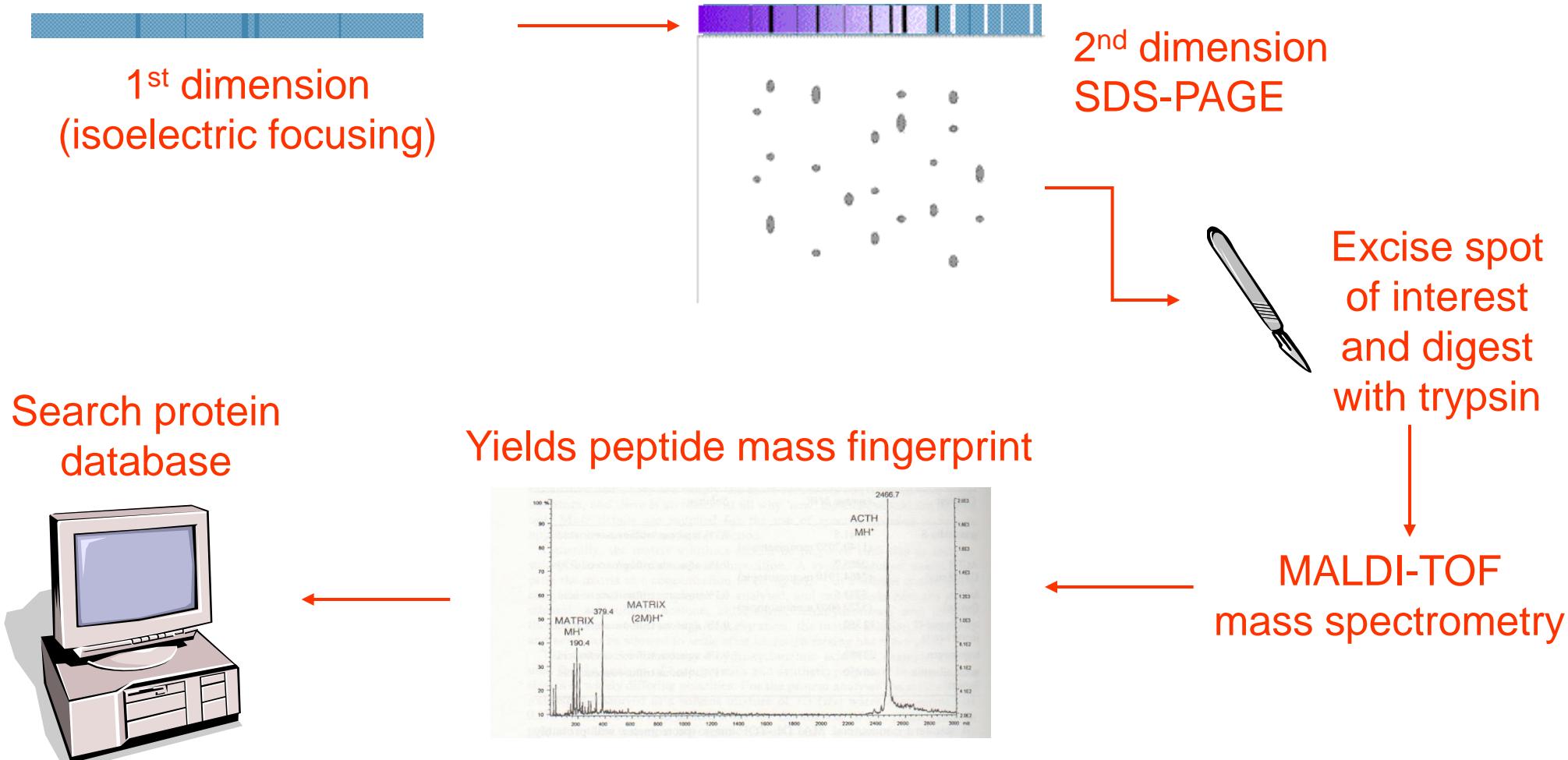
University of Glasgow



- **Proteomics and milk proteins**
 - Gel based proteomics
 - Non-gel based proteomics
- **Metabolomics and mastitis**
- **Non-invasive sampling**
 - Saliva
 - Sweat
 - Nasal secretion
 - Interstitial fluid



2-DE and mass spectrometry



- **Galvani et al Rapid Comm in Mass Spec, 2000, 14:1889**
- **Spots:**
 - 9-11; β -lactoglobulin
 - 13-15; α -lactalbumin
 - Others; caseins

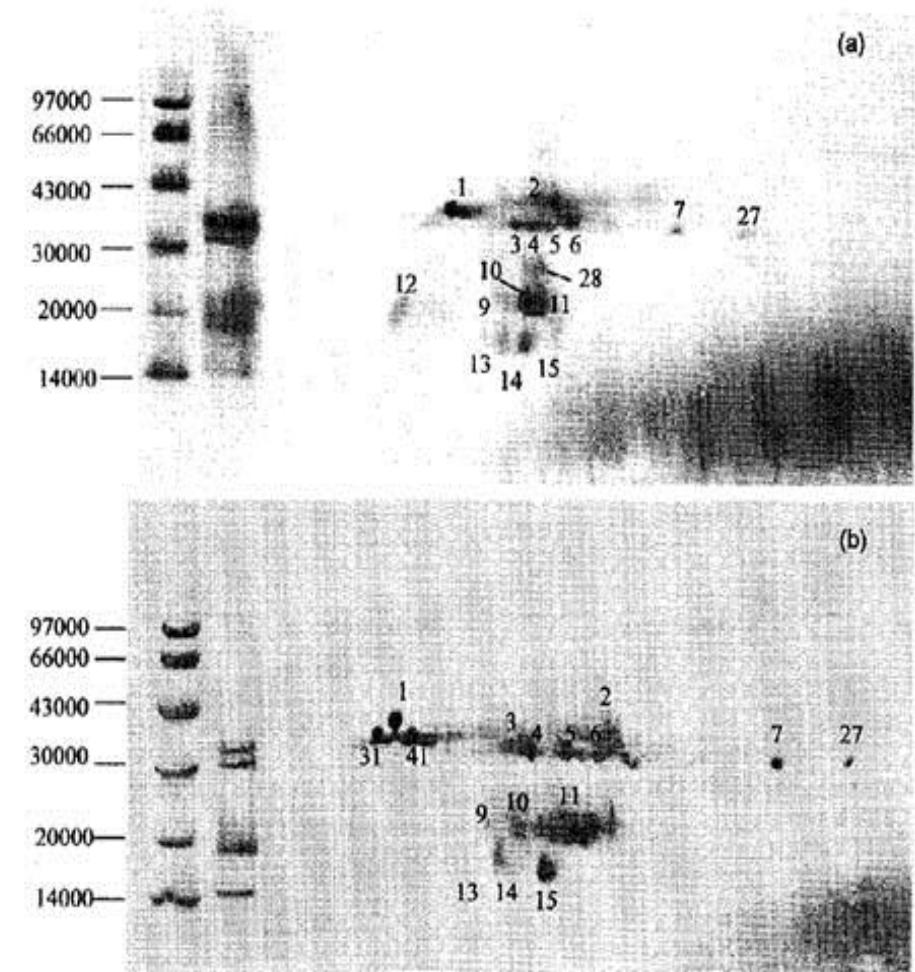
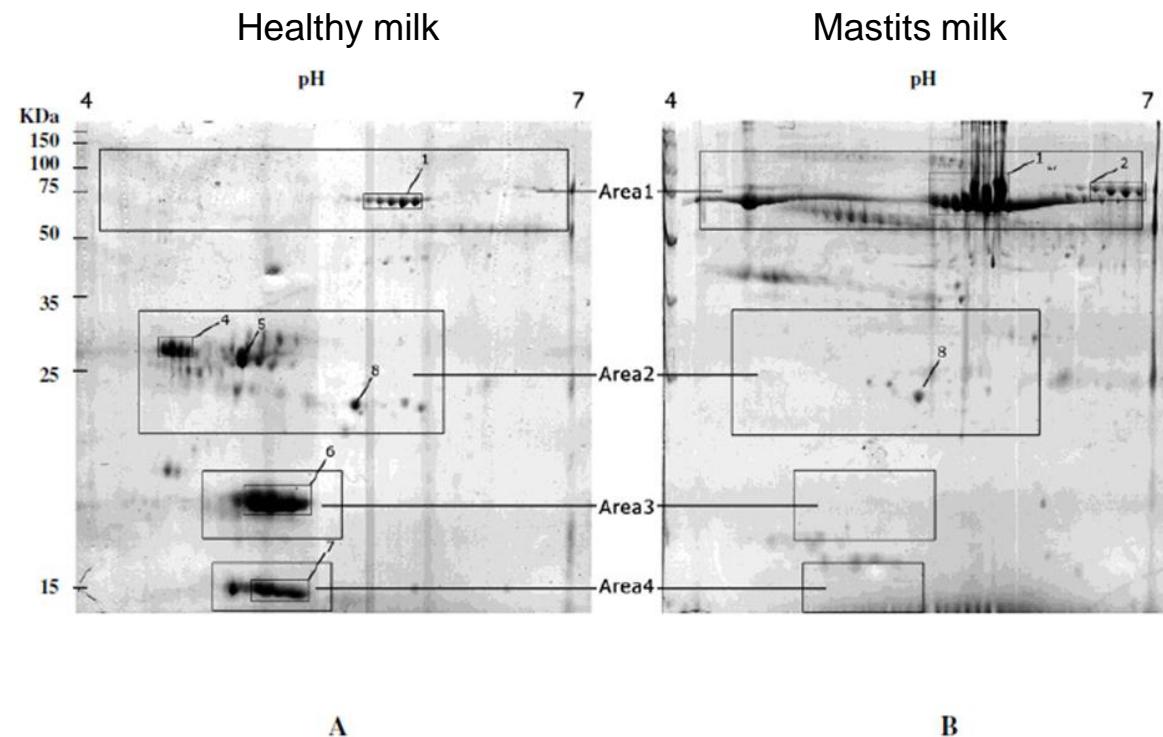


Figure 2. 2-D gel maps stained with Coomassie Brilliant Blue: (a) pH gradient 3–7 and (b) pH gradient 4–7.

Milk in clinical mastitis

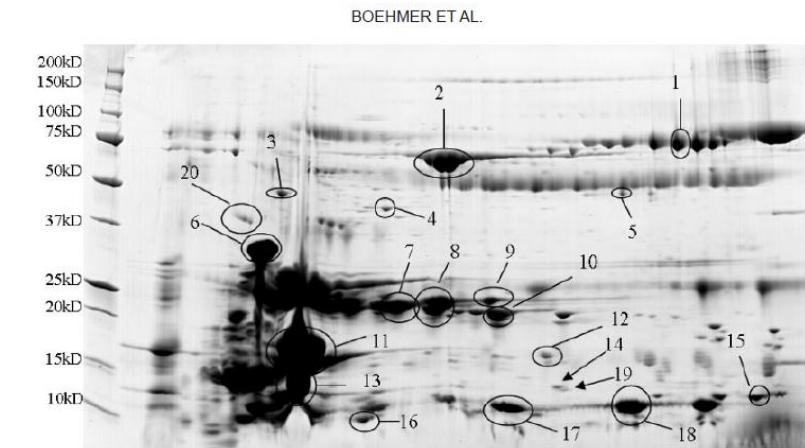
Increased: albumin,
lactoferrin (area 1), IgG

Decreased: casein (area 2),
 β -lactoglobulin (area 3),
 α -lactalbumin (area 4)



Hogarth et al 2004

- Boehmer et al 2008
- Ultracentrifugation to prepare samples
- High abundance of casein and beta-lactoglobulin,
- Healthy whey samples show low abundance proteins: transthyretin, lactadherin, beta-2-microglobulin, alpha-1-acid glycoprotein, and complement C3



- Mastitis whey samples increased serum albumin, transthyretin, complement C3 and alpha-1-acid glycoprotein and appearance of cathelicidin, indolicidin, and bactenecin 5 and 7, beta-fibrinogen, alpha-2-HS-glycoprotein, alpha-1-antiproteinase

- **2 Dimension electrophoresis based proteomics**

- Separate proteins on polyacrylamide gel
- Stain and pick protein spots
- Trypsin digest → peptides
- Mass spectrometry → peptide mass
- Protein identification by bioinformatic interrogation

- **2 Dimension electrophoresis based proteomics**

- Separate proteins on polyacrylamide gel
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- Trypsin digest → peptides
- Mass spectrometry → peptide mass
- Protein identification by bioinformatic interrogation

- **Non-gel proteomics**

- Trypsin digest → peptides OR ultrafiltration to remove large protein
- Include labelled standards for quantification
- Separate peptides on Liquid Chromatography or Capillary Electrophoresis
- Mass spectrometry → peptide mass
- Protein identification by bioinformatic interrogation
- Calibrate with labelled standards

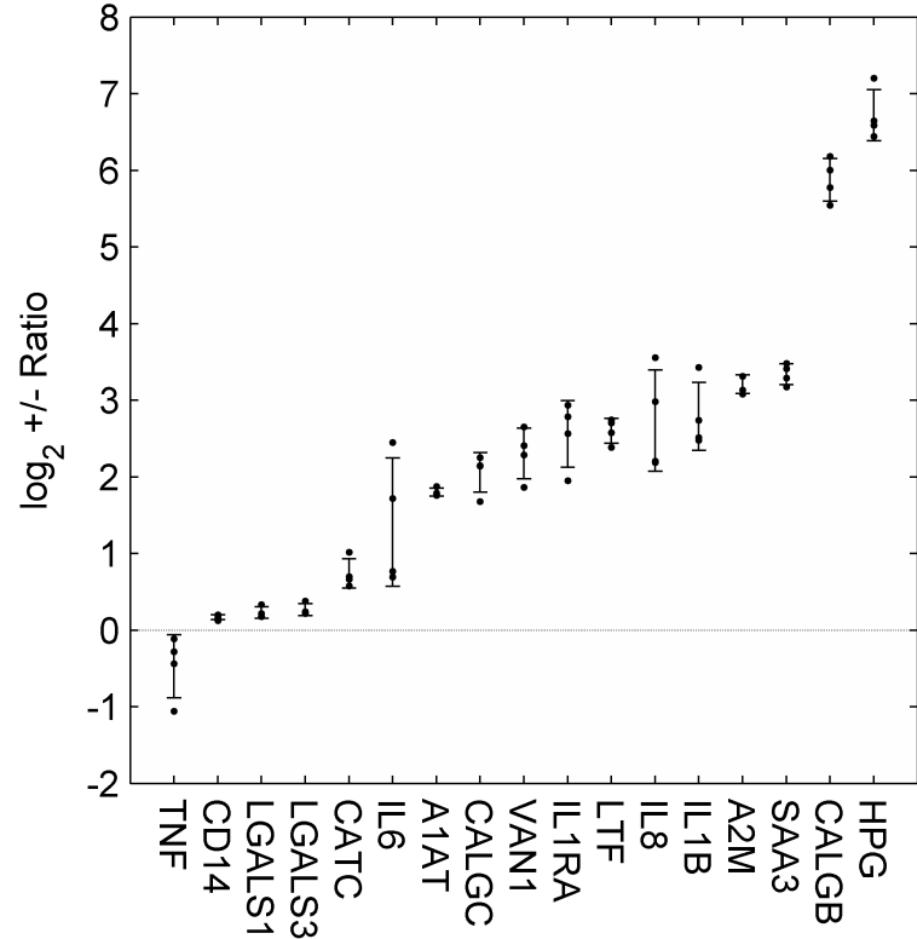
- **Danielson et al 2010 Proteomics 10:2240**
- **iTRAQ and LC-MS/MS**
- **Quantitative analysis of 80 milk proteins**
- **49 proteins changed in milk following experimental mastitis with *E. coli* infection of udder**
- **Increase compared to controls:**
 - Apolipoprotein I, II & IV
 - Bactenecins 5 & 7
 - Complement C3 and C4

- Reinhardt et al J Proteomics (2013) 82:141
- Proteomes determined in milk exosomes, milk fat globule membranes (MFGM) & whey
- iTRAQ & LC-MS/MS
- >300 milk proteins identified related to host defence
- 94 significantly altered in *Staphylococcus aureus* infection of udder
- Identified neutrophil extracellular trap formation in MFGM

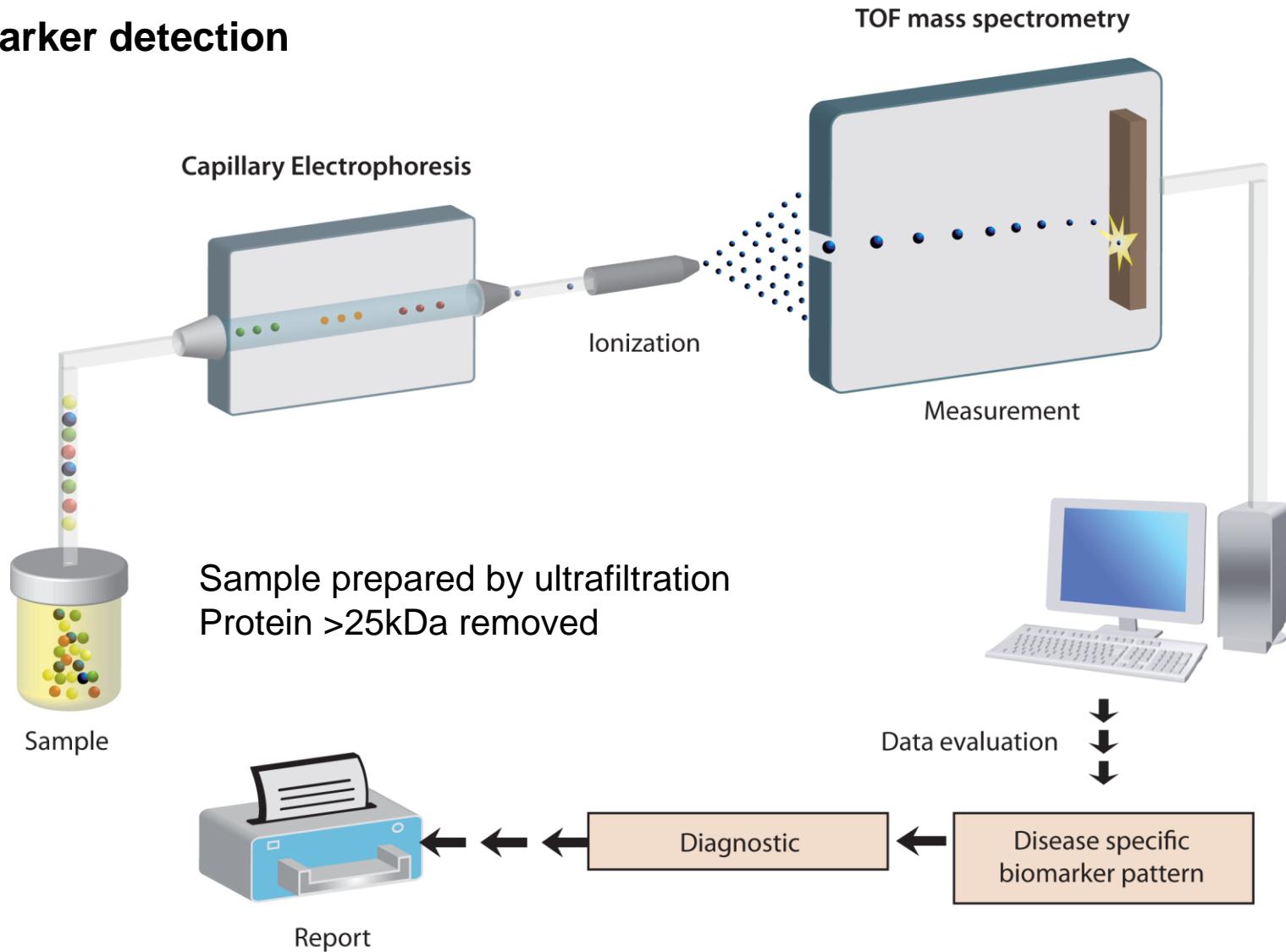
Quantotypic Properties of QconCAT Peptides Targeting Bovine Host Response to *Streptococcus uberis*

Stine L. Bislev,^{†,‡} Ulrike Kusebauch,^{‡,§} Marius C. M. Rønnow,[‡] Christine M. Røntved,[†] Ruedi Aebersold,^{#,¶,△,||}

J Prot Res, 2012, 11:1832



Biomarker detection



Control (n=12) and clinical mastitis milk (n=25)

Clinical mastitis detected by swelling, heat, pain, clots in milk

Ultrafiltered to remove protein >25kDa

Run on CE-MS

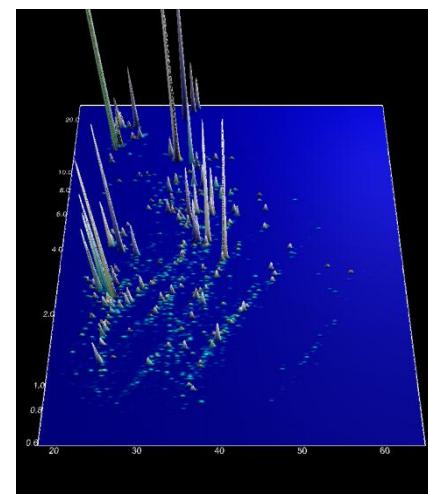
Peaks combined by statistical analysis to develop algorithm

>1000 peptide peaks detected

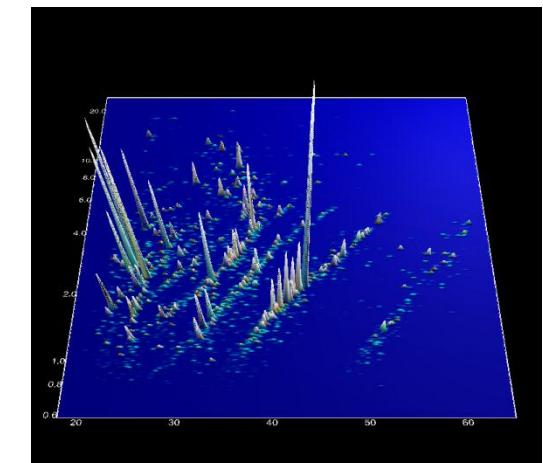
508 peptides differ between healthy & mastitic milk

Mansor et al *Journal of Proteomics* 2013 85:89

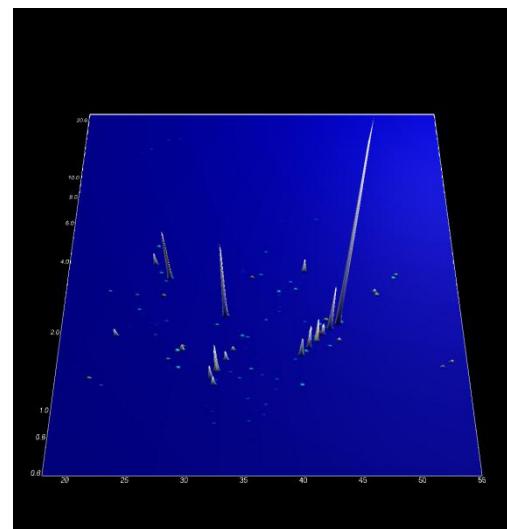
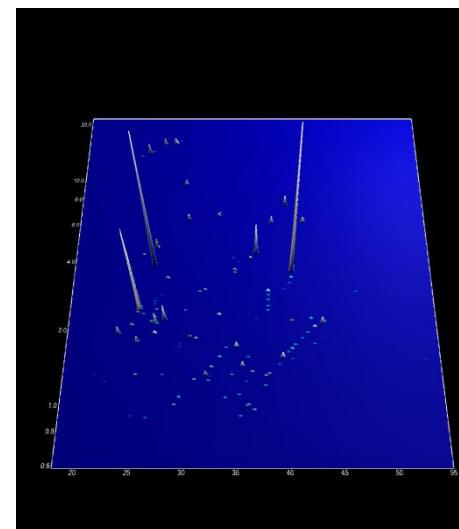
Healthy



Mastitis



Greater in healthy



Greater in mastitis

Peaks combined by statistical analysis

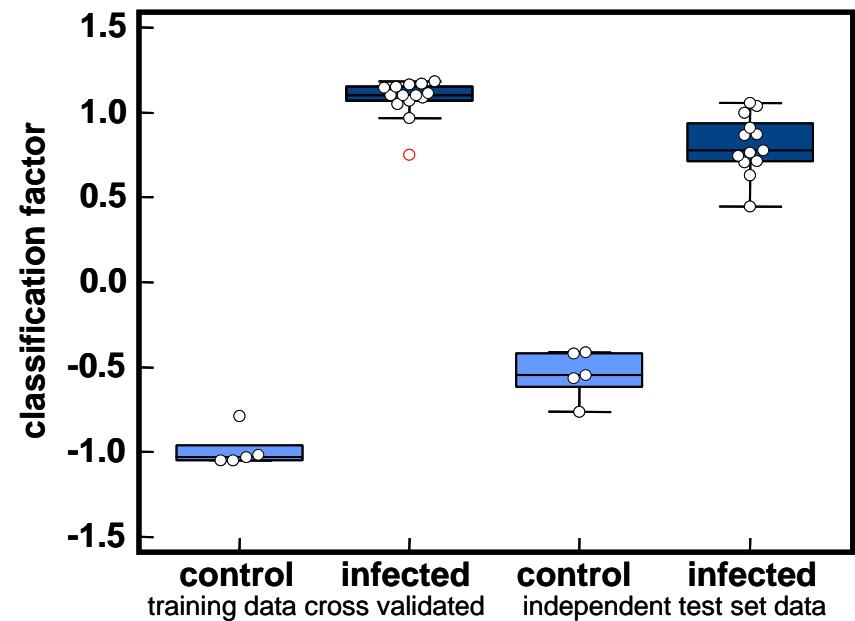
154 peptide multiplex biomarker panel

Training sets: 5 healthy samples v 14 clinical mastitis samples

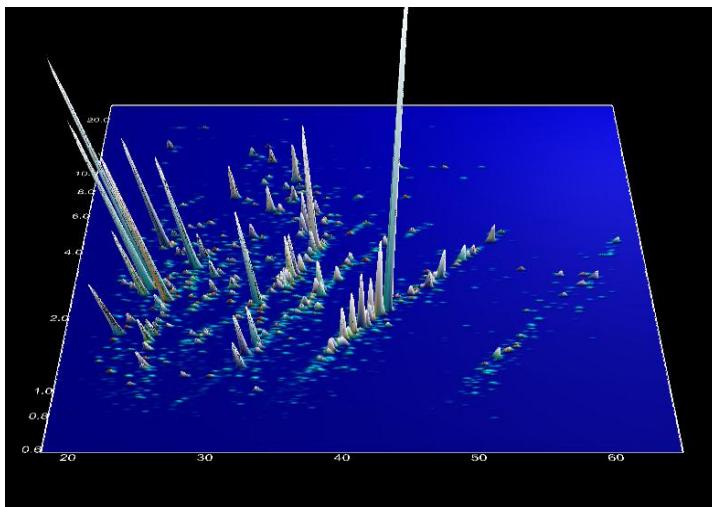
Test sets: 5 healthy samples v 13 clinical mastitis samples

100% sensitive & 100% specific

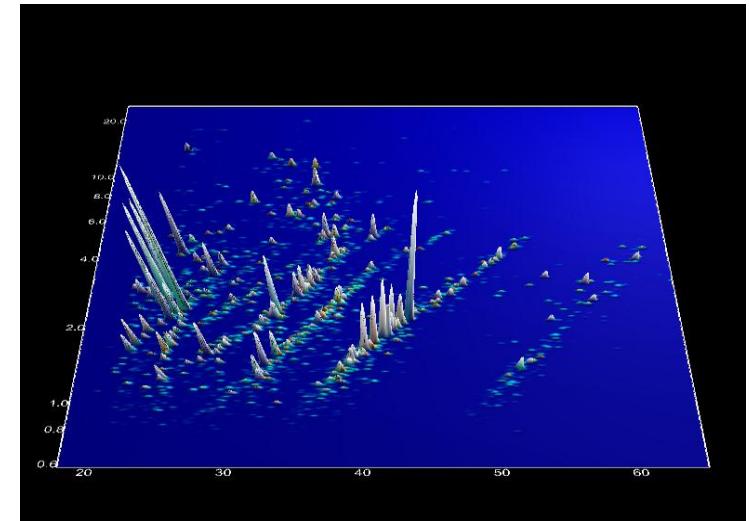
Peptides identified mostly casein breakdown products



Differential diagnosis of bacterial cause of mastitis



E. Coli



S. aureus

Bacterial infection determined by Glasgow Veterinary Diagnostic Service

Peaks combined by statistical analysis (Mosaique Diagnostics)

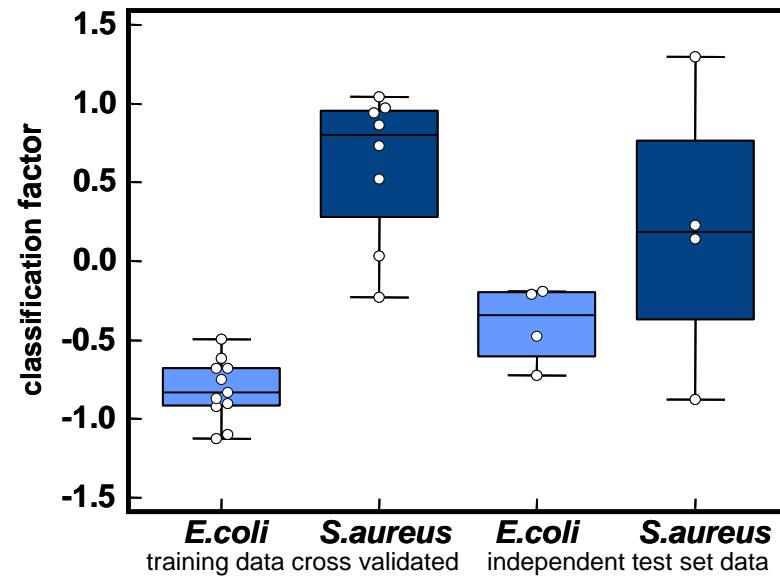
47 peptide multiplex biomarker panel

Training sets: 11 *E. Coli* v 8 *S. aureus* samples

Test sets: 4 *E. Coli* v 4 *S. aureus* samples

75% sensitive & 100% specific

Peptides identified mostly casein breakdown products



Metabolomics

- Characterization and quantification of the whole collection of both intra and extra-cellular metabolites.

Metabolome

- the qualitative and quantitative collection of all low molecular weight of molecules
- in biological fluid, organism or cell
- can be peptides, amino acids, nucleic acids, carbohydrates, organic acids, vitamins, polyphenols, alkaloids

- Does the milk metabolome change with mastitis?
- Are there differences in the milk metabolome with different pathogens?
- Can change in specific metabolites be used as biomarkers for mastitis?

Mass spectrometry analysis

- Milk extracted with methanol:chloroform:water (3:1:1)
- LC system coupled with MS (Dionex UltiMate 1)
- MS Acquisition (Orbitrap Exactive)
- Performed in both –ve and +ve ionisation modes and full scan modes

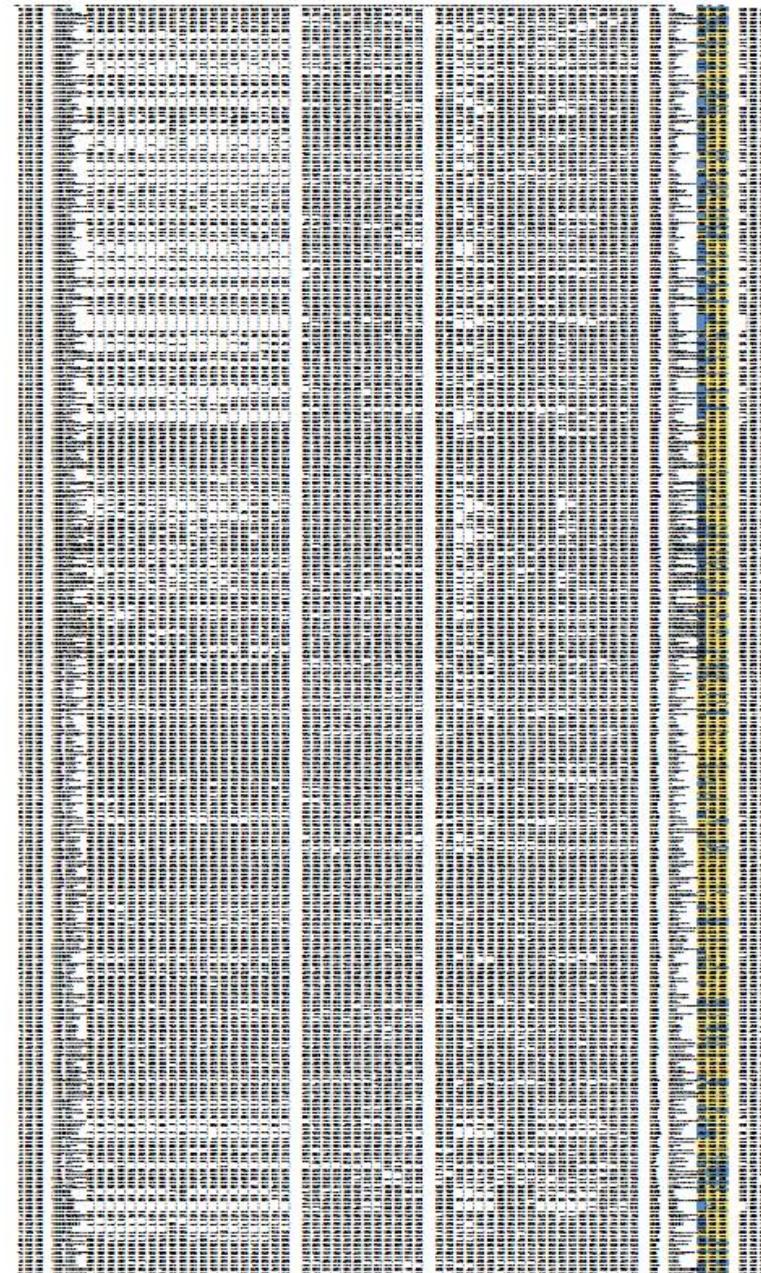
- 10 Healthy milk samples
- 10 *E.Coli* milk samples
- 6 *Staph. aureus* milk samples

- 5502 MS negative ion peaks
- 573 identified metabolites

- 9908 MS positive ion peaks
- 814 identified metabolites

- 1 of 12 pages of MS data for negative ion data set

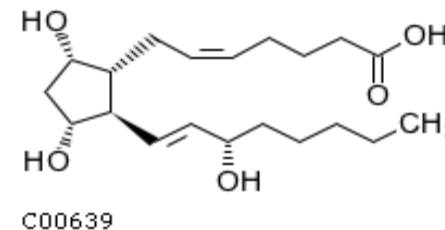
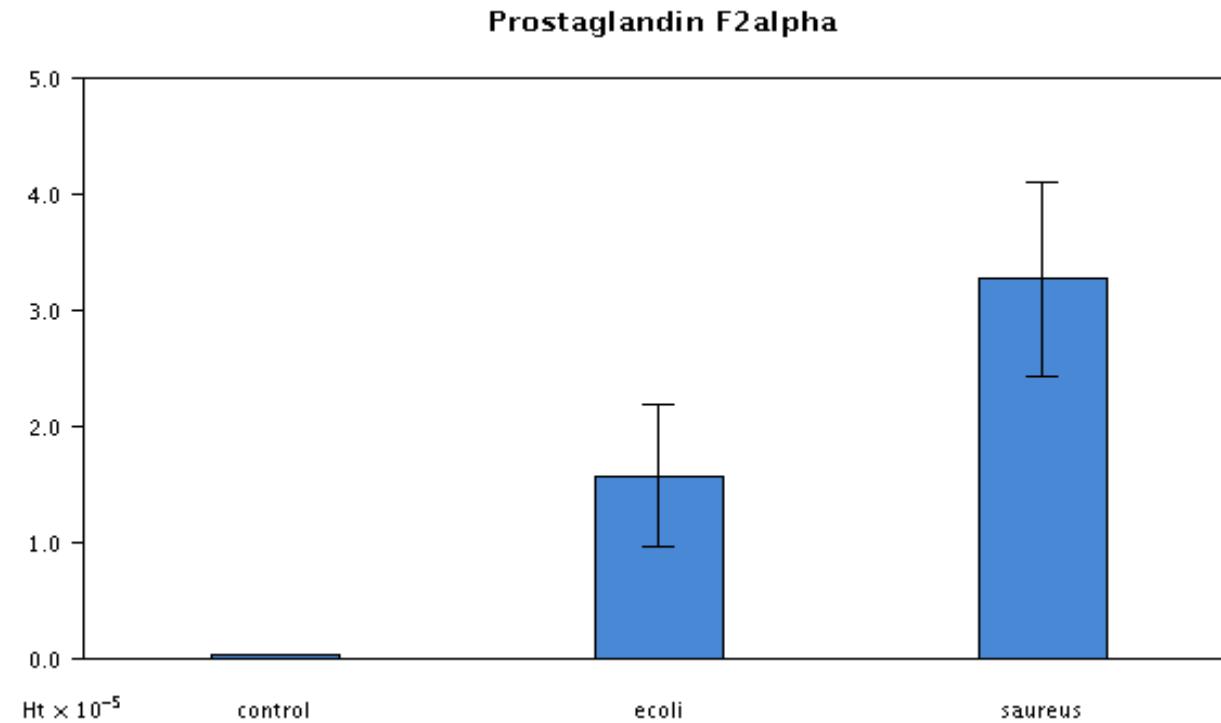
- Challenge for bioinformatics



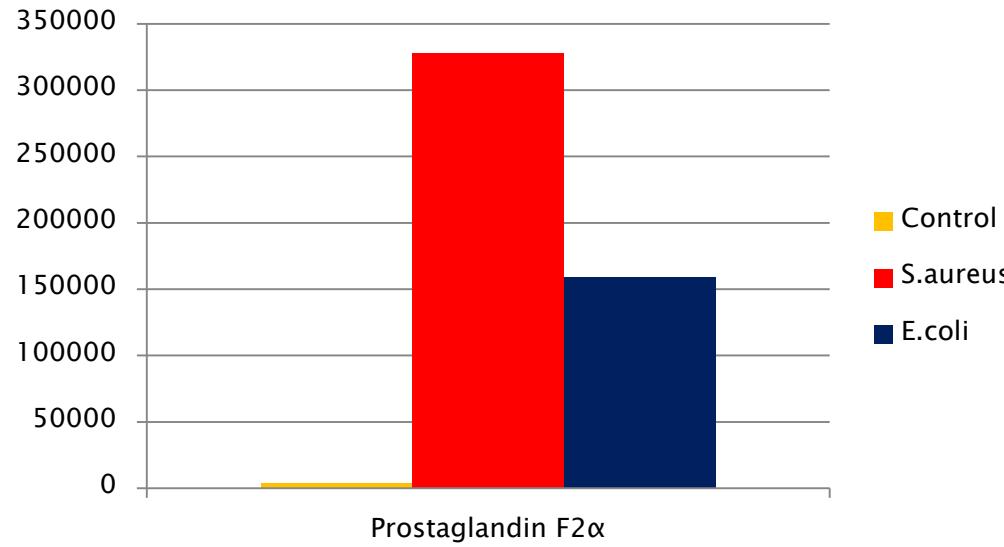
The first 50 of 573 peaks (Negative Ion Mode)

| Metabolites | Control | Name | E.Coli | Name | S.aureus |
|-------------------------------------|----------|---------------------------------------|----------|---------------------------------------|----------|
| 5-Dehydro-4-deoxy-D-glucarate | 1.75E+09 | 5-Dehydro-4-deoxy-D-glucarate | 1.38E+09 | 5-Dehydro-4-deoxy-D-glucarate | 1.71E+09 |
| 1-alpha-D-Galactosyl-myo-inositol | 1.51E+09 | 1-alpha-D-Galactosyl-myo-inositol | 1.16E+09 | 1-alpha-D-Galactosyl-myo-inositol | 1.25E+09 |
| N-Acetyl-D-glucosamine 6-phosphate | 2.62E+08 | (R)-Lactate | 8.12E+08 | (R)-Lactate | 9.22E+08 |
| cis-Aconitate | 1.67E+08 | myo-Inositol | 1.83E+08 | L-Glutamate | 1.56E+08 |
| myo-Inositol | 1.06E+08 | Succinate | 1.8E+08 | cis-Aconitate | 1.24E+08 |
| Hippurate | 98680533 | cis-Aconitate | 1.05E+08 | myo-Inositol | 1.16E+08 |
| Urate | 89863389 | 2-C-Methyl-D-erythritol 4-phosphate | 91564904 | (S)-Malate | 1E+08 |
| 2,5-Dioxopentanoate | 84341727 | 2-Dehydro-3-deoxy-L-arabinonate | 86395991 | Urate | 94020844 |
| Miserotoxin | 78910119 | Pyruvate | 81948401 | 2-C-Methyl-D-erythritol 4-phosphate | 85004997 |
| 2-C-Methyl-D-erythritol 4-phosphate | 78482788 | Creatine | 73380634 | L-Erythulose | 74942006 |
| L-Erythulose | 75351299 | Hippurate | 72131854 | N-Acetyl-D-glucosamine | 73804064 |
| N-Acetyl-D-glucosamine | 74489145 | (S)-Malate | 71188741 | Miserotoxin | 70788430 |
| Creatine | 74407132 | 2-C-Methyl-D-erythritol 4-phosphate | 63327089 | 2,5-Dioxopentanoate | 69476176 |
| (S)-Malate | 73918031 | Urate | 60860379 | Taurine | 62376397 |
| L-Glutamate | 73661846 | L-Glutamate | 60367636 | (R)-3-Hydroxybutanoate | 62310337 |
| sn-glycero-3-Phospho-1-inositol | 66333040 | 2,5-Dioxopentanoate | 59941285 | Pyruvate | 58442163 |
| D-Glucose 6-phosphate | 52176193 | N-Acetyl-D-glucosamine 6-phosphate | 59306858 | Creatine | 56614851 |
| 2-Dehydro-3-deoxy-L-arabinonate | 44090003 | Glycerol 2-phosphate | 51930002 | Hippurate | 5370833 |
| 2-Butyne-1,4-diol | 43558519 | 2-Oxoglutarate | 47839083 | 2-Dehydro-3-deoxy-L-arabinonate | 45261272 |
| Pantothenate | 43488200 | (2S)-2-Isopropylmalate | 46199319 | Mesaconate | 42718964 |
| 2-Oxoglutarate | 41739044 | Taurine | 43331718 | 2-Butyne-1,4-diol | 38833822 |
| Glycerophosphoglycerol | 35372625 | N-Acetyl-D-glucosamine | 37618824 | N-Acetyl-D-glucosamine 6-phosphate | 35648796 |
| L-Alanine | 32778314 | Pantothenate | 36966410 | Succinate | 29519867 |
| 2-Octadecanoic acid | 31016415 | L-Erythulose | 32796067 | L-Alanine | 25713006 |
| (R)-Lactate | 30676971 | Mesaconate | 32519002 | Pantothenate | 25649695 |
| 2-C-Methyl-D-erythritol 4-phosphate | 17129722 | Miserotoxin | 32238333 | L-Arginine | 24661840 |
| Phenylacetylglycine | 16394154 | 2-Butyne-1,4-diol | 31756211 | Acetoacetate | 23241854 |
| Glycerol 2-phosphate | 15741286 | L-Alanine | 30500954 | Leucyl-leucine | 23052455 |
| Uridine | 15169547 | D-Glycerate | 21083181 | (R)-2-Hydroxystearate | 21361935 |
| D-Glycerate | 13282239 | (R)-3-Hydroxybutanoate | 20058371 | L-1-Pyrroline-3-hydroxy-5-carboxylate | 21140417 |
| 2-Hydroxy-2,4-pentadienoate | 11126302 | Glycerophosphoglycerol | 17011504 | L-Aspartate | 20924711 |
| Fumarate | 10123233 | alpha-D-Mannoheptulopyranose | 14571959 | Glycerophosphoglycerol | 20292854 |
| 1-Aminocyclopropane-1-carboxylate | 8933048 | Glycerophosphoglycerol | 14066463 | Leu-Gln-Ser | 18858881 |
| Acetoacetate | 8114826 | Diacyl | 13189653 | Lys-Val-Ser | 18012351 |
| Pyruvate | 7995273 | L-Arginine | 12673627 | 2-Oxoglutarate | 17828323 |
| Succinate | 5936534 | (S)-3-Methyl-2-oxopentanoic acid | 12014074 | Diacyl | 17016063 |
| L-2-Amino adipate | 5451488 | D-Glucose 6-phosphate | 10716111 | 2-Octadecanoic acid | 16754393 |
| (S)-Dihydroorotate | 5164245 | Fumarate | 9768704 | Leu-Tyr | 15228717 |
| D-Glucuronolactone | 5130855 | sn-glycero-3-Phospho-1-inositol | 9457991 | D-Glycerate | 15214056 |
| Taurine | 5106576 | Leu-Tyr | 8427266 | Leu-Thr | 14873123 |
| Met-Asn-Asp | 5096341 | 2-Octadecanoic acid | 8162199 | Fumarate | 14530951 |
| Xyitol | 5046532 | Met-Asn-Asp | 8105485 | Leu-Phe | 13157998 |
| Fumarate | 4844560 | (R)-2-Hydroxystearate | 7952443 | Leu-Leu-Val | 12690777 |
| 1-4-beta-D-Glucan | 4819930 | Phenylacetyl-glycine | 7817072 | 2-C-Methyl-D-erythritol 4-phosphate | 12648423 |
| (R)-3-Hydroxybutanoate | 4219629 | L-1-Pyrroline-3-hydroxy-5-carboxylate | 7631977 | 9-10-DHOME | 11837381 |
| alpha-D-Mannoheptulopyranose | 3797943 | Acetoacetate | 7415715 | Uridine | 11031207 |
| Bicine | 3590543 | 1-4-beta-D-Glucan | 6210871 | Leu-Trp | 10963377 |
| 4-Oxoproline | 3581575 | Fumarate | 6149763 | Val-Arg | 10020592 |
| 4-Pyridoxate | 3464541 | 5-Hydroxypentanoate | 5744097 | 1-Aminocyclopropane-1-carboxylate | 9663711 |

Change in individual metabolites

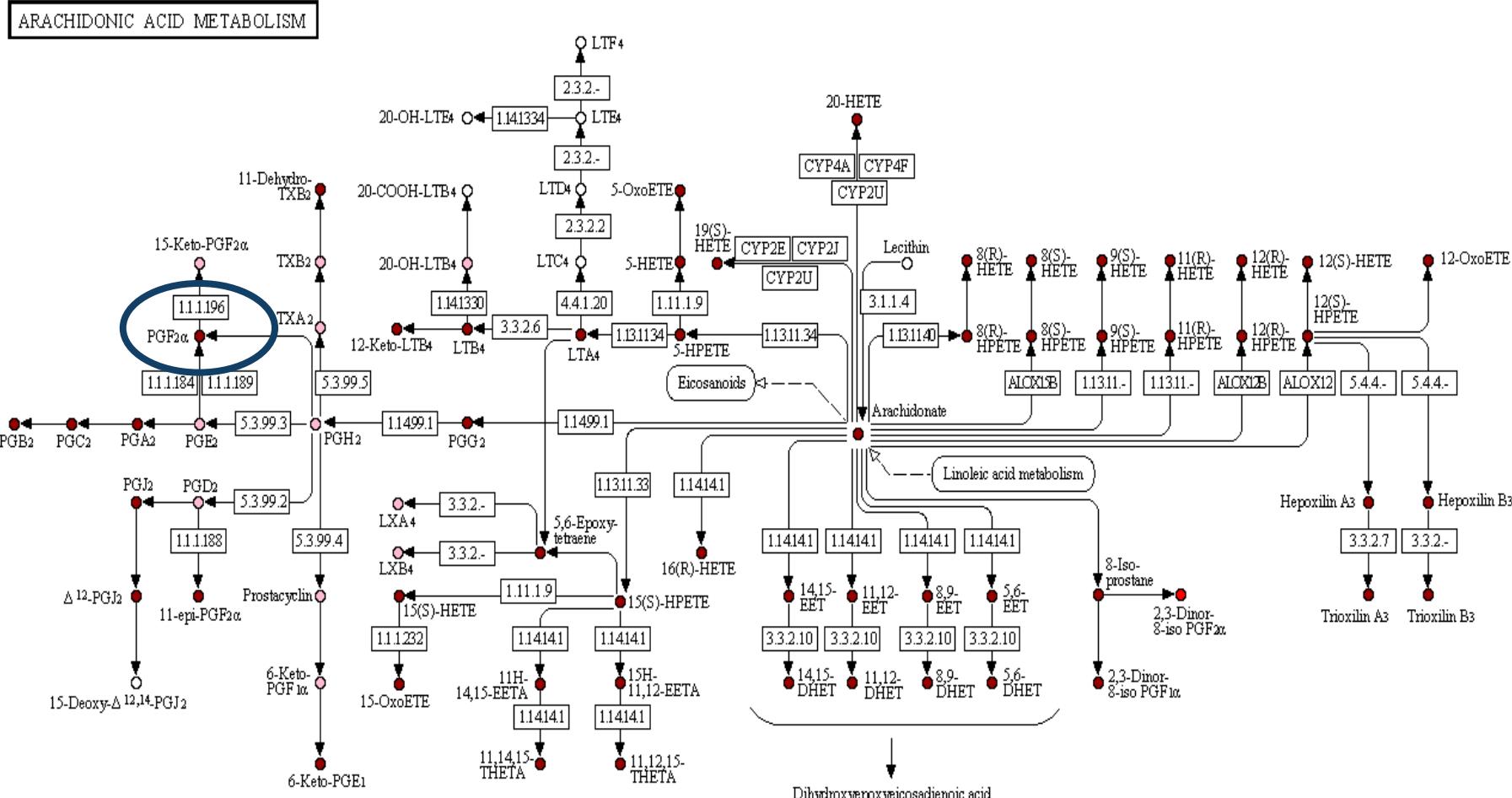


Kegg map metabolites with formula $C_{20}H_{34}O_5$

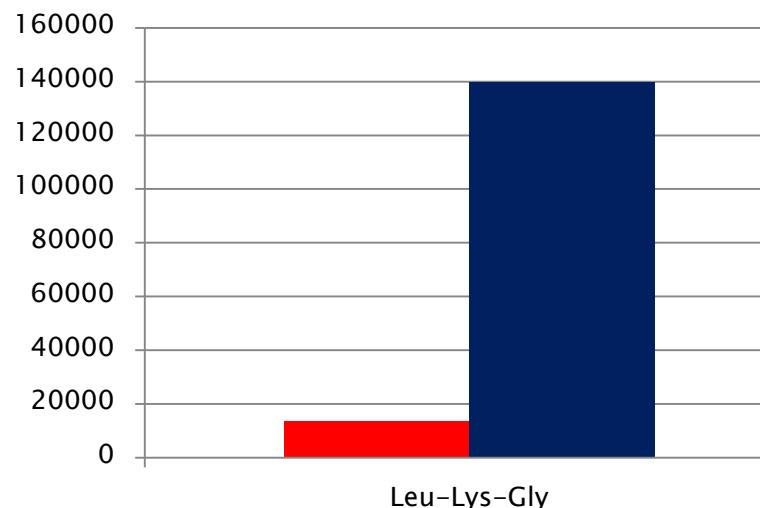
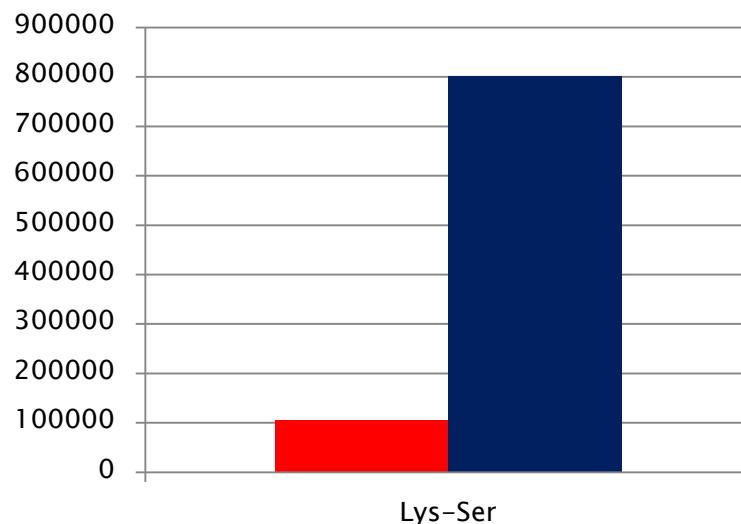
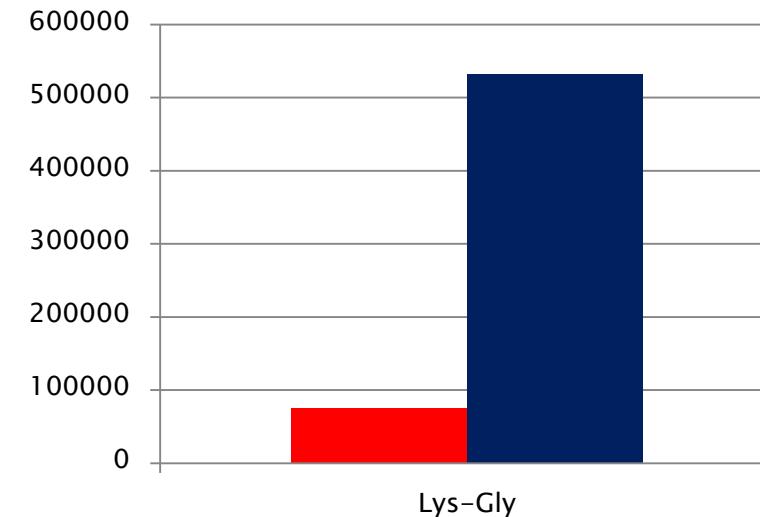
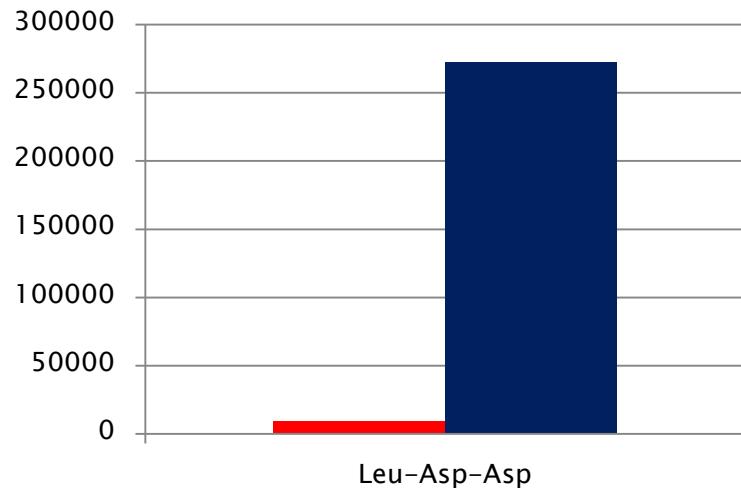


Prostaglandin F 2α are significantly higher in both bacterial groups compared with control group

Kegg Diagram of pathway changes: red dot increased in mastitis

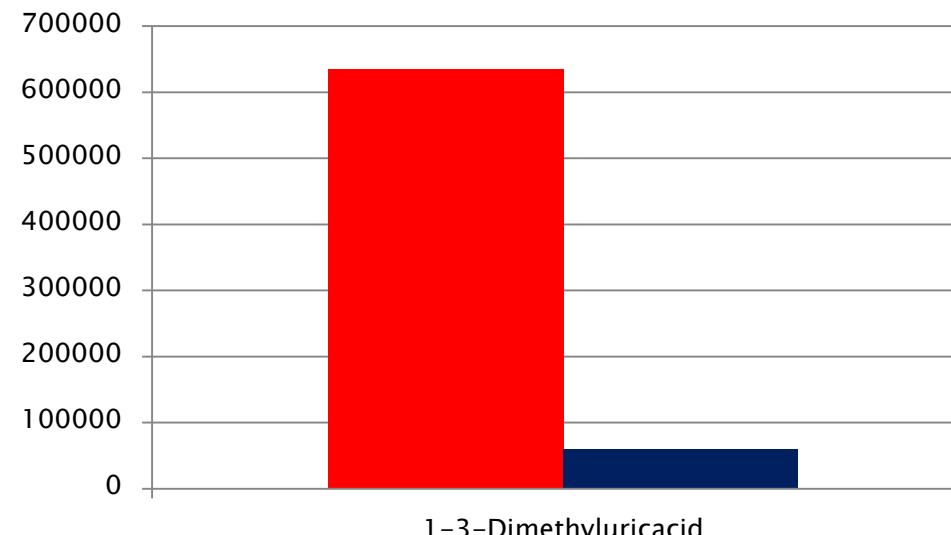
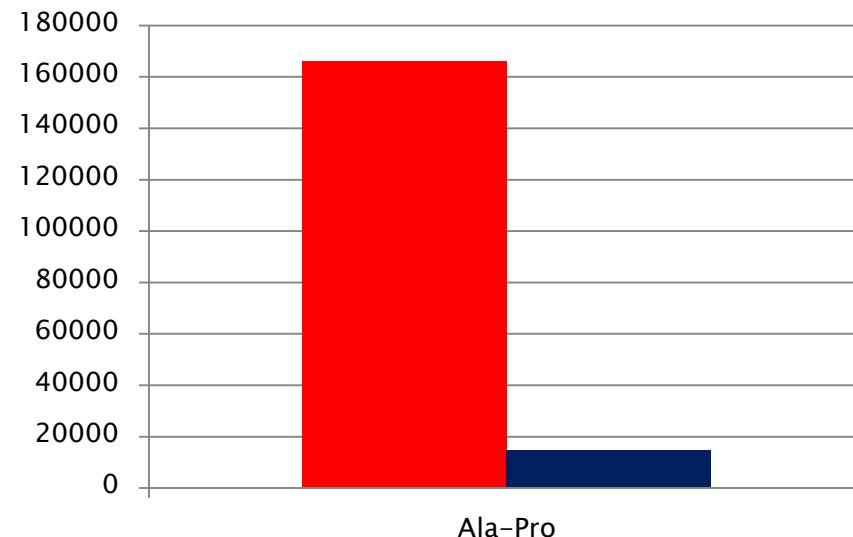
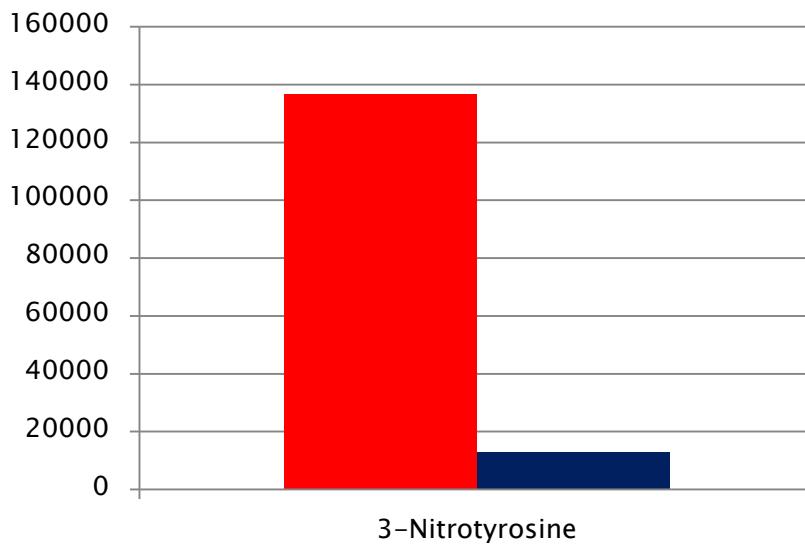


Metabolites found highest in *S. aureus* group



 *S.aureus*
 *E.coli*

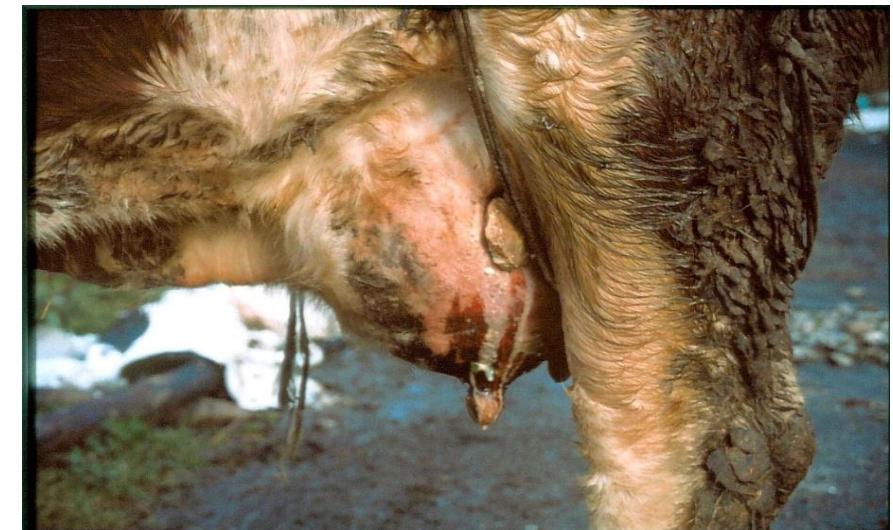
Metabolites found highest in *E.coli* group



 *S. aureus*
 *E. coli*

- Lipid, protein and carbohydrate metabolism are all affected by mastitis
- Arachidonic acid metabolites are increased in mastitis
- Amino acids, di & tri peptides increase in mastitis
- Significant metabolite differences found in healthy versus mastitis milk
- Significant metabolite differences found in *E. coli* versus *S. aureus* milk
- Many other causes of change in milk metabolome

- **Proteomics and milk proteins**
 - Gel based proteomics
 - Non-gel based proteomics
- **Metabolomics and mastitis**
- **Non-invasive sampling**
 - Saliva
 - Sweat
 - Nasal secretion
 - Interstitial fluid



- Bovine saliva collected on gauze swabs
- 4 different pre-fractionation methods
- 402 salivary protein
- 45 N-linked glycoprotein
- Immunoglobulin, prolactin-inducible protein, cystatins, carbonic anhydrase, lactoferrin, lysozyme
- No amylase
- Ang et al J Prot Res 2011 10 5059

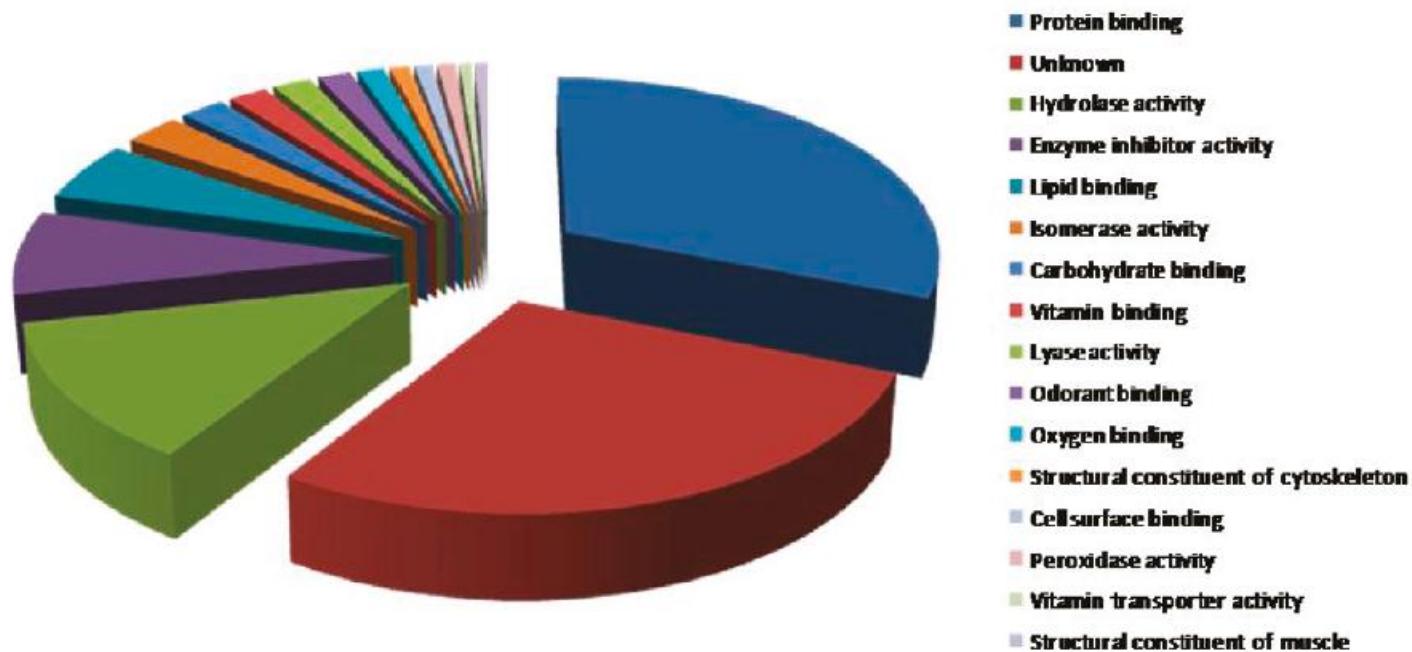


Figure 4. Functional classification of the 396 proteins identified from the nontargeted approach (SDS-PAGE, Off-gel, RP-HPLC and SCX-HPLC prefractionation). Classification is based on the GeneOntology molecular function annotation using the DAVID Functional Annotation Clustering tool (<http://david.abcc.ncifcrf.gov/>).

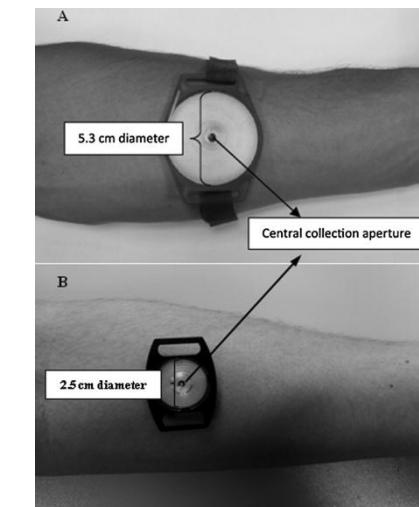
- Human sweat proteome
- Armpit sweat compared in controls v Schizophrenia patients
- LC-MS/MS analysis
- 17 protein had 2x or higher difference in patient group
- Raiszadeh et al J Prot Res 2012, 11:2127

| Protein | Peptide | Patient Pool: Control Pool |
|------------------------------|--------------------|----------------------------|
| Alpha-2-glycoprotein | YSLTYIYTGLSK | 1.9 : 1 |
| | AYLEEECPATLR | 1.7 : 1 |
| Annexin 5 | VLTEIIASR | 4.6 : 1 |
| | GTVTDFPGFDER | 5.1 : 1 |
| Arginase | GGVVEEGPTVLR | 2.9 : 1 |
| Bleomycin Hydrolase | DGEAVWFGCDVGK | inf. |
| Calmodulin-like Skin Protein | VNYEEFAR | 2.1 : 1 |
| | AGLEDLQVAFR | 2.0 : 1 |
| Caspase 14 | FQQAIDSR | 2.7 : 1 |
| | LENLFEALNNK | 5.4 : 1 |
| Corneodesmosin | IILQPCGSK | 2.5 : 1 |
| | GSPGVPSFAAGPPISEGK | 3.0 : 1 |
| Cystatin A | TQVVAGTNYYIK | 2.8 : 1 |
| Dermcidin | ENAGEDPGLAR | 1.0 : 1 |
| Desmoglein | YQGTILSIDDNLQR | 3.6 : 1 |
| DJ-1 | DGLILTSR | 4.0 : 1 |
| G3PDH | VPTANVSVDLTCR | 6.7 : 1 |
| Kallikrein 11 | LLCGATLIAPR | 2.6 : 1 |
| Keratin 10 | YENEVALR | 1.0 : 1 |
| | ALEESNYELEGK | 0.97 : 1 |
| Peroxiredoxin 1 | GLFIIDDK | 6.2 : 1 |
| | LVQAFQFTDK | 9.3 : 1 |
| Prolactin-induced Protein | TVQIAAVVDVIR | 0.57 : 1 |
| Prostatic-binding Protein | LYTLVLTDPDAPSR | 7.2 : 1 |
| S100A7 | GTNYLADVFEK | 9.2 : 1 |
| Thioredoxin | VGEFSGANK | 8.9 : 1 |
| | TAFQEALDAAGDK | 10.6 : 1 |

- **Sweat collecting device required 10µl for metabolomics**

- **Compared sweat from control and lung cancer patient**

- **Possible biomarkers:**
citrulline, maltotriose, maltotetraose, suberic, urocanic, muconic acid and nonanedioic acids, trisaccharide phosphate, phenylalanine, histidine, taurine, citruline, tryptophan, sulfonic lipid, and l-octen-3yl-glucoside



- Human sweat examined by LC-MS/MS

- Antimicrobial found in sweat

- Lactoferrin confirmed by western blot

Park et al Exper Derm 2011 20 367

Table 1. AMP identified by Mass spectrometry from human sweat sample

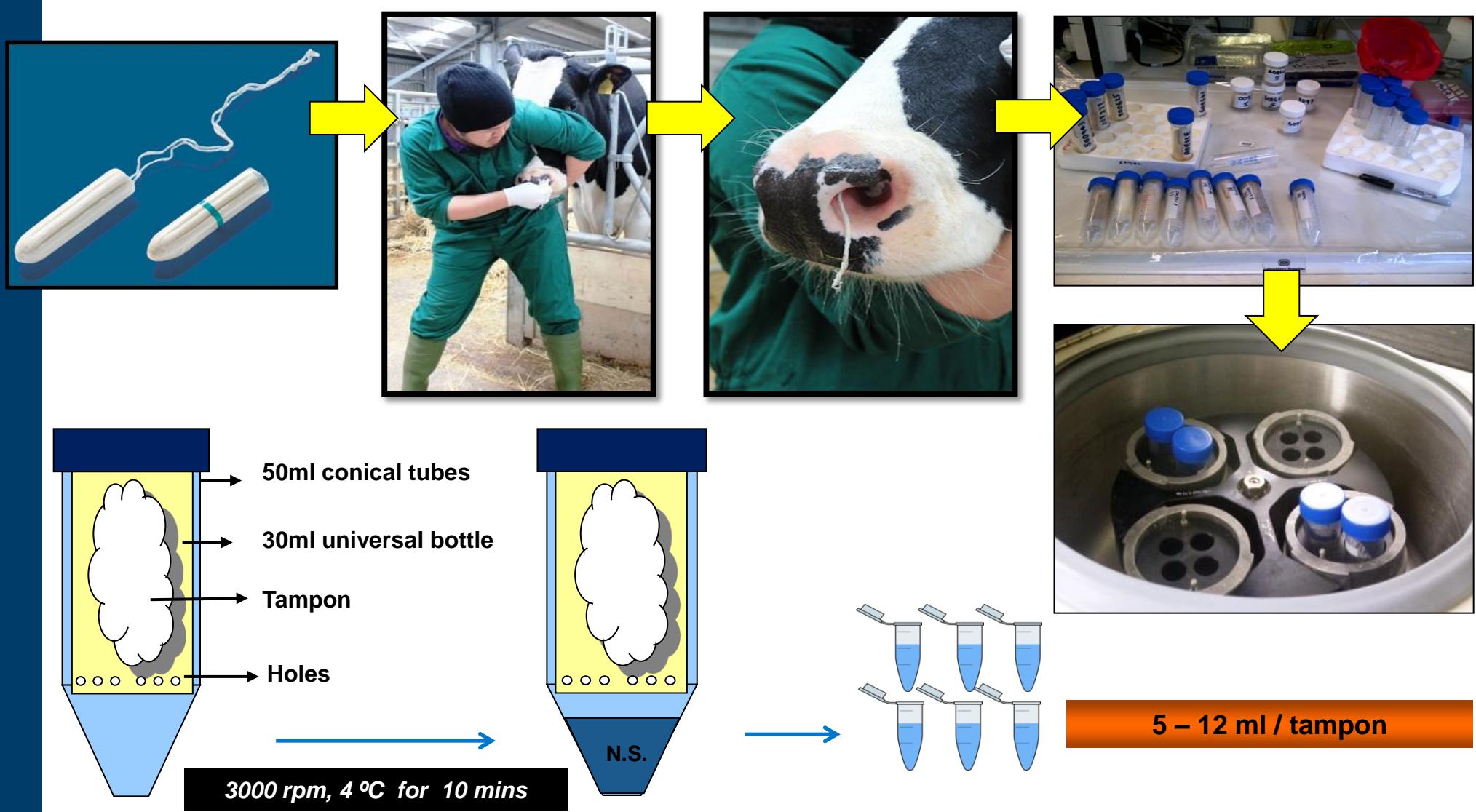
| Accession number | Mass [Da] | Protein score | Description |
|------------------|-----------|---------------|---|
| gi 34412 | 78 029 | 828 | Lactoferrin |
| gi 16751921 | 11 277 | 299 | Dermcidin precursor |
| gi 12053626 | 10 410 | 238 | Psoriasin |
| gi 299033 | 14 679 | 128 | Lysozyme = amyloid fibril protein [human, Peptide Mutant, 130 aa] |
| gi 31979 | 13 899 | 107 | Histone H2A.2 |
| gi 7245541 | 37 076 | 75 | Chain a, r210k n-terminal lobe human lactoferrin |
| gi 13399625 | 14 678 | 71 | Chain A, Mutant Human Lysozyme (q86d) |
| gi 307141 | 16 544 | 70 | Lysozyme precursor (EC 3.2.1.17) |
| gi 386772 | 15 238 | 48 | Histone H3 |
| gi 32097 | 11 269 | 47 | Histone cluster 2, h4a |

AMP, antimicrobial peptides or proteins.

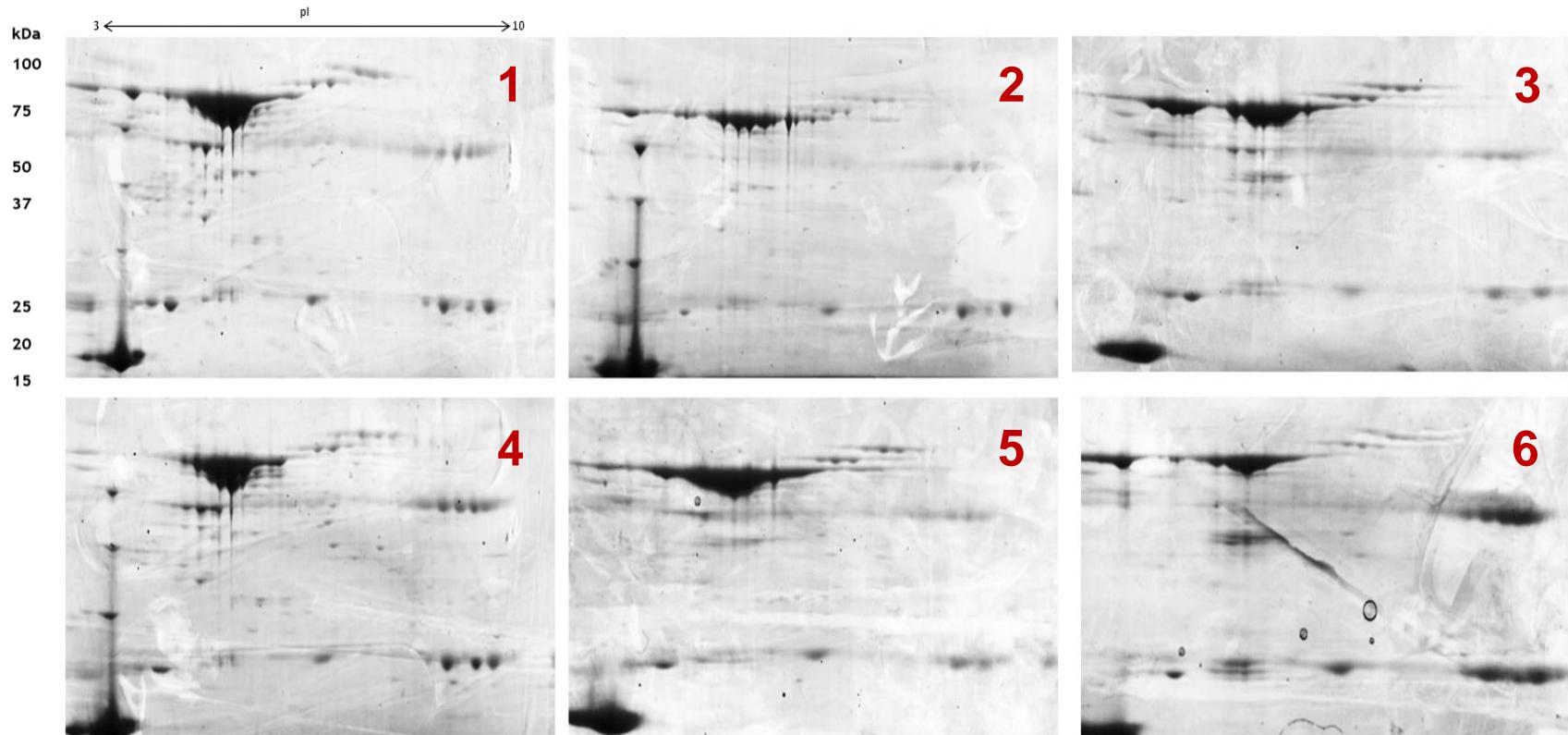
| Proteome list from sweat by LC-MS/MS | | | | |
|--------------------------------------|------------------|-----------|---------------|---|
| | Accession number | Mass [Da] | Protein score | Description |
| 1 | gi 1346343 | 65978 | 989 | Cytokeratin 1 |
| 2 | gi 34412 | 78029 | 828 | Lactoferrin |
| 3 | gi 29667 | 53117 | 781 | Carboxypeptidase E |
| 4 | gi 38026 | 34714 | 779 | Zn-alpha2-glycoprotein |
| 5 | gi 28317 | 59492 | 668 | Cytokeratin 10 |
| 6 | gi 619383 | 27975 | 578 | Apolipoprotein D |
| 7 | gi 435476 | 62092 | 484 | Cytokeratin 9 |
| 8 | gi 56554584 | 17914 | 473 | Chain A, Crystal Structure Of Human Tear lipocalin von EBNERS GLAND Protein |
| 9 | gi 42716297 | 57796 | 400 | Clusterin isoform 1 |
| 10 | gi 229532 | 8446 | 388 | Ubiquitin |
| 11 | gi 178855 | 48772 | 379 | Apolipoprotein J precursor |
| 12 | gi 55859601 | 15883 | 345 | Calmodulin-like 5 |
| 13 | gi 51094526 | 16562 | 335 | Prolactin-induced protein |
| 14 | gi 6912286 | 27662 | 333 | Caspase 14 precursor |
| 15 | gi 5902072 | 44537 | 319 | Serine (or cysteine) proteinase inhibitor, clade B (ovalbumin), member 3 |
| 16 | gi 4557014 | 59719 | 299 | Catalase |
| 17 | gi 16751921 | 11277 | 299 | Dermcidin precursor |
| 18 | gi 57997569 | 52459 | 266 | Hypothetical protein |
| 19 | gi 483474 | 65289 | 264 | 90K , galectin 3 binding protein |
| 20 | gi 28590 | 69250 | 246 | Albumin |
| 21 | gi 4504963 | 19238 | 238 | Lipocalin 1 precursor |
| 22 | gi 12053626 | 10410 | 238 | Psoriasin |
| 23 | gi 238236 | 83262 | 232 | Transmembrane secretory component; poly-Ig receptor; SC |
| 24 | gi 189308 | 53698 | 229 | Nucleobindin |
| 25 | gi 5729909 | 9918 | 224 | Lipophilin B precursor |

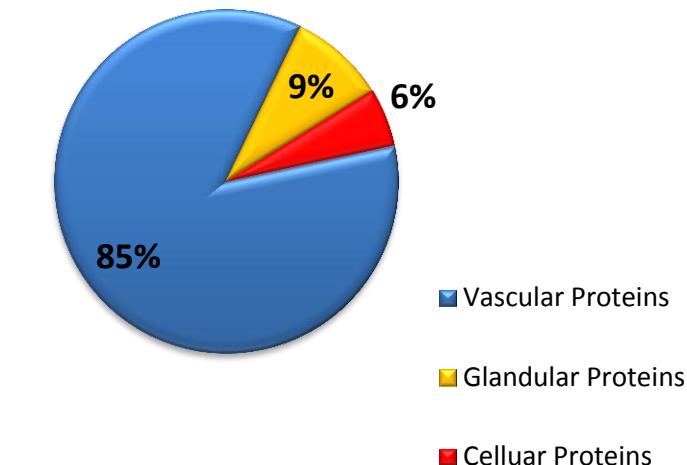
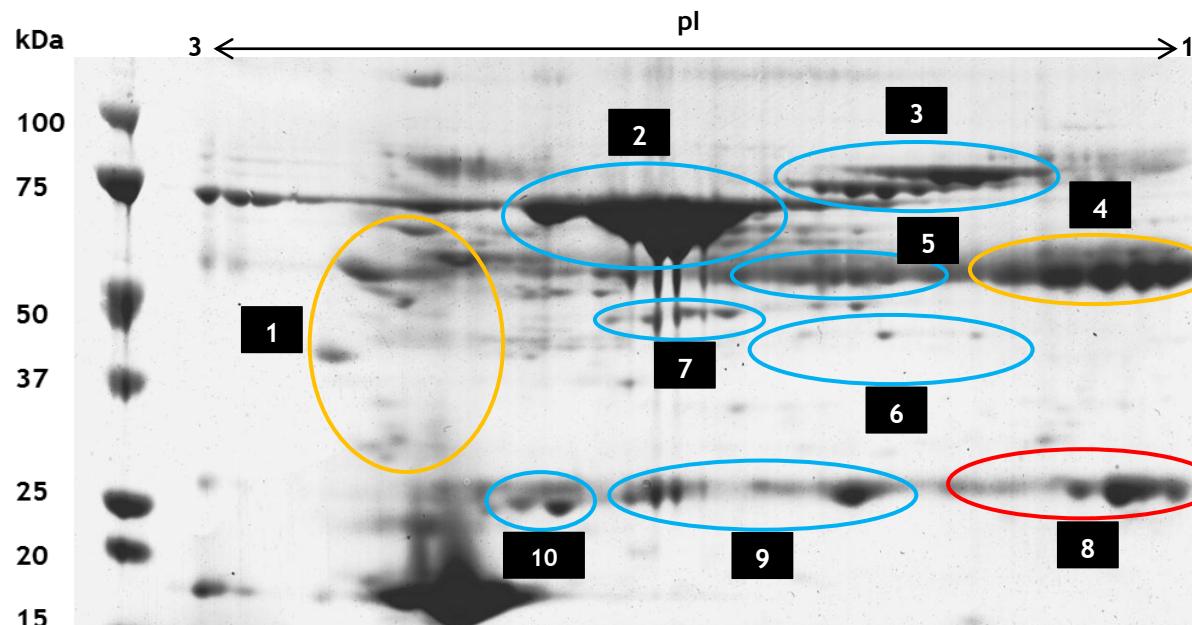
- Nasal secretion for non-invasive sampling
 - Nasal secretion collection – up to 12ml in 10 min
 - Biochemical analysis revealed high activity of alkaline phosphatase
 - Protein content examined by 2DE
 - Nasal secretion proteome characterised
 - Change in proteins in response to malignant catarrhal fever determined
-
- Mohd Faizal Ghazali, University of Glasgow

Nasal secretion - Collection



Individual 2-D electrophoresis of nasal secretion
from six different cattle

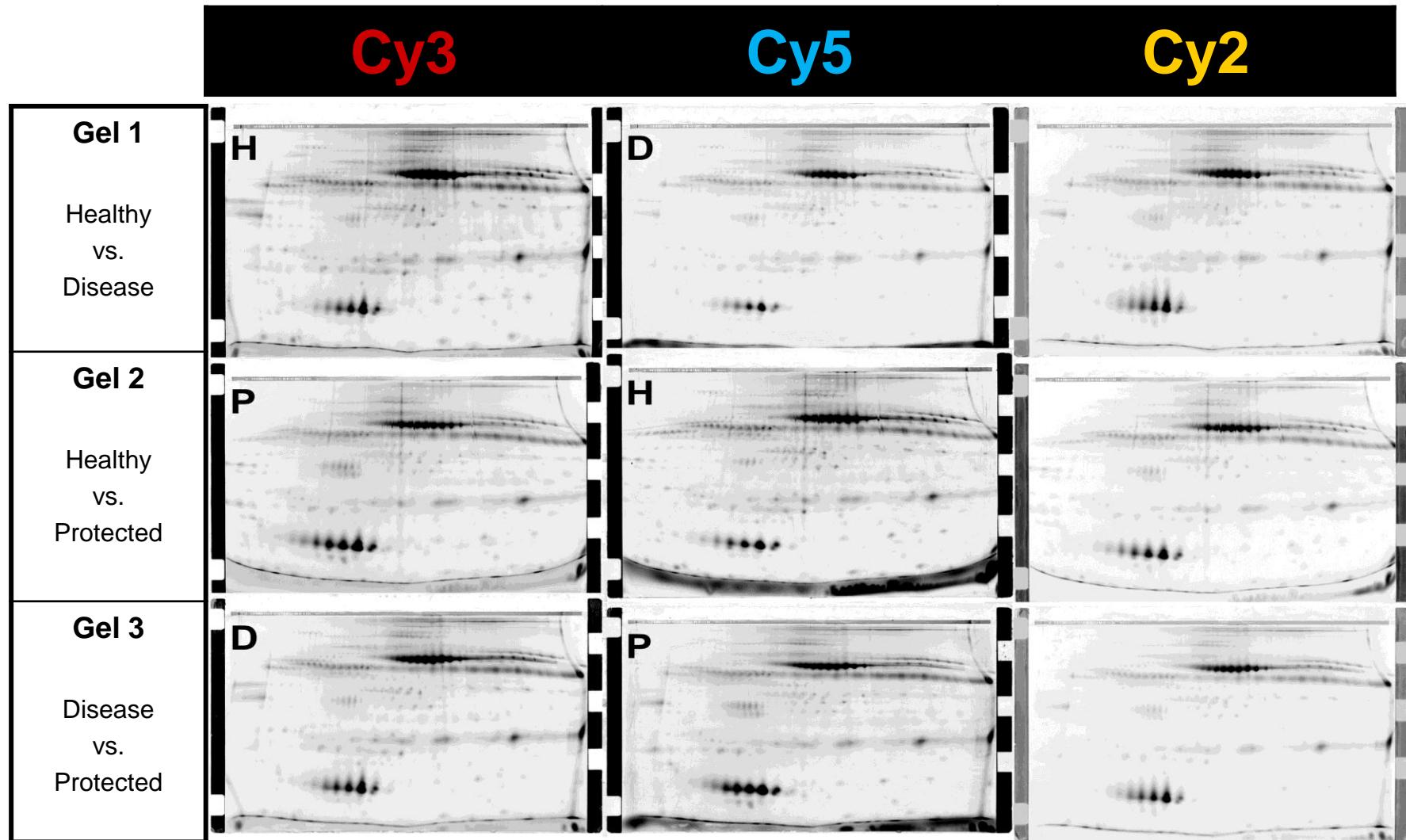




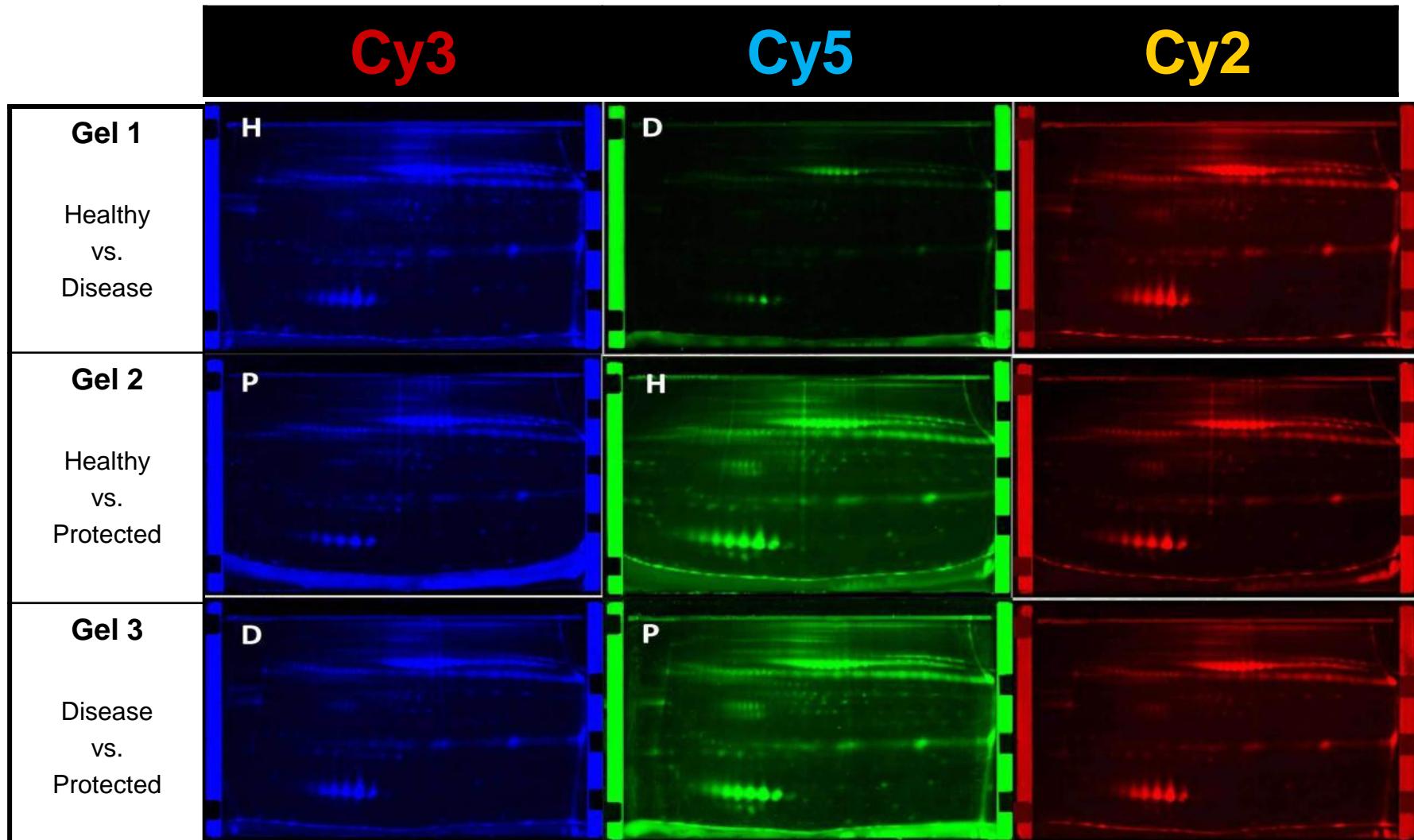
| Protein spot no. | Protein name | Accession | Theoretical mass (Da) | Highest MS score | Highest Matches |
|------------------|---------------------------|--------------|-----------------------|------------------|-----------------|
| 1 | Odorant-binding protein | OBP_BOVIN | 18492 | 498 | 32 |
| 2 | Albumin | ALBU_BOVIN | 71244 | 2568 | 163 |
| 3 | Serotransferrin | TRFE_BOVIN | 79870 | 1394 | 102 |
| 4 | Lactotransferrin | TRFL_BOVIN | 80002 | 129 | 12 |
| 5 | Ig heavy chain | O46780_BOVIN | 51391 | 621 | 38 |
| 6 | Fibrinogen beta chain | FIBB_BOVIN | 53933 | 474 | 40 |
| 7 | Complement C3 | CO3_BOVIN | 188652 | 501 | 51 |
| 8 | Glutathione S-transferase | GSTP1_BOVIN | 23826 | 525 | 26 |
| 9 | Ig light chain | Q1RMN8_BOVIN | 25032 | 189 | 23 |
| 10 | Apolipoprotein A1 | APOA1_BOVIN | 30258 | 1296 | 77 |

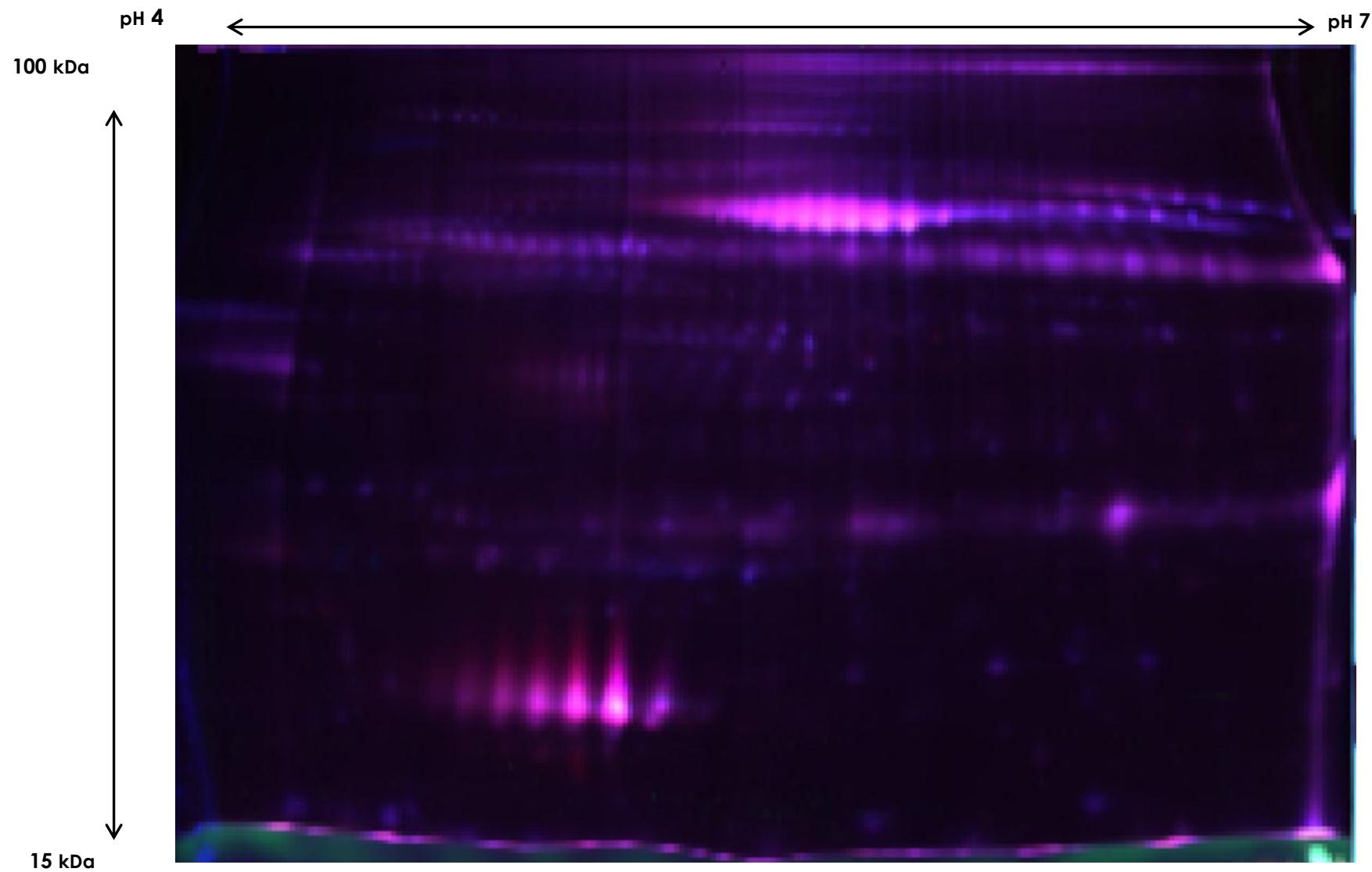
- Nasal secretion from experimental model of Malignant Catarrhal Fever
- Calves immunised with attenuated C500 strain AIHV-1
- Challenge with virulent C500 strain AIHV-1
- Healthy samples: Pre-challenge NS sample
- Disease: not immunised, NS taken at 2 weeks, no survivors at week 12
- Protected: immunised, NS taken at 2 weeks, all survived at week 12
- Pools of NS from each group prepared (n=7) and run on DiGE in duplicate

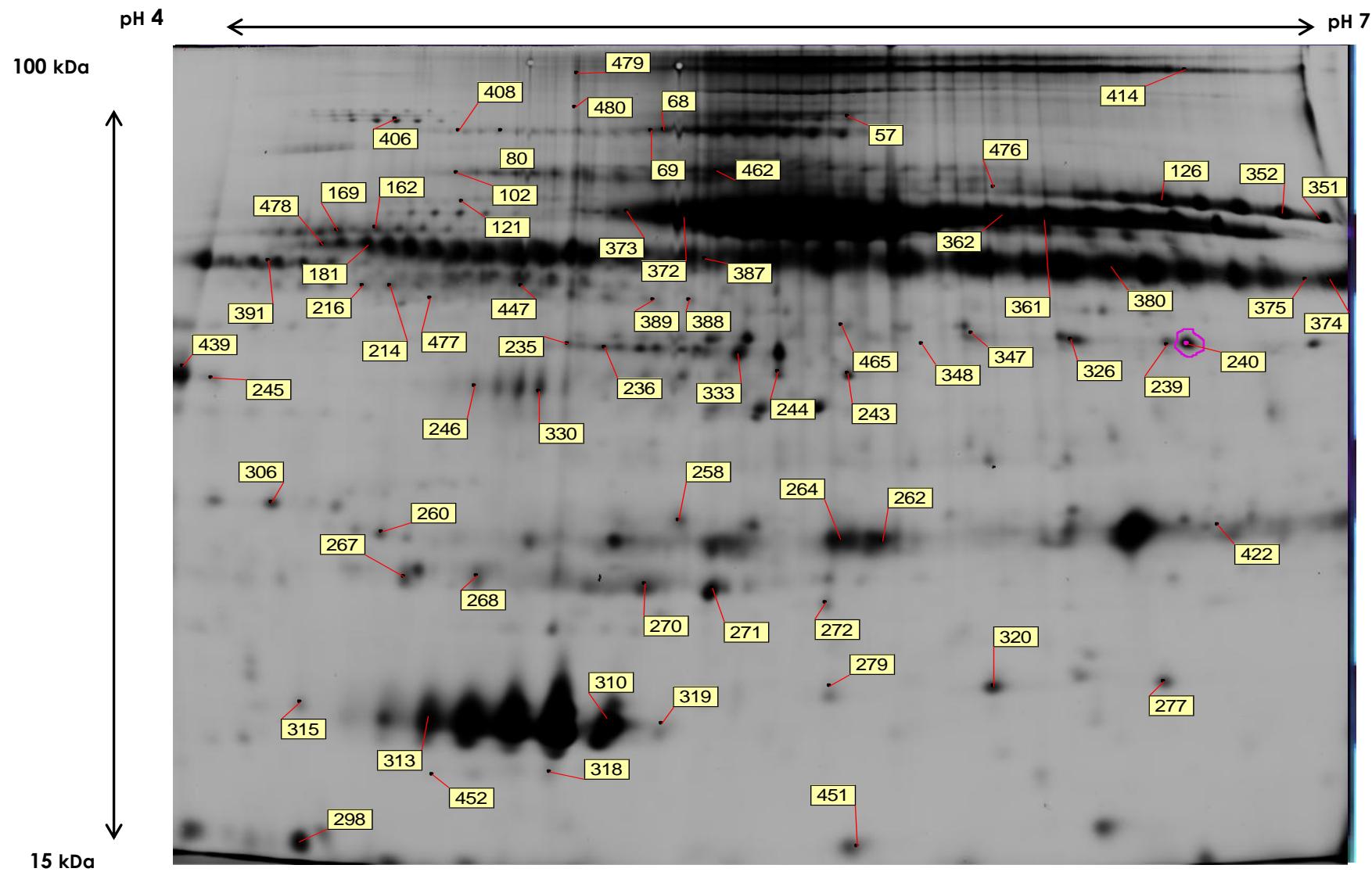
Relative Quantification by Difference Gel Electrophoresis (DiGE) of Nasal secretion from calves with MCF



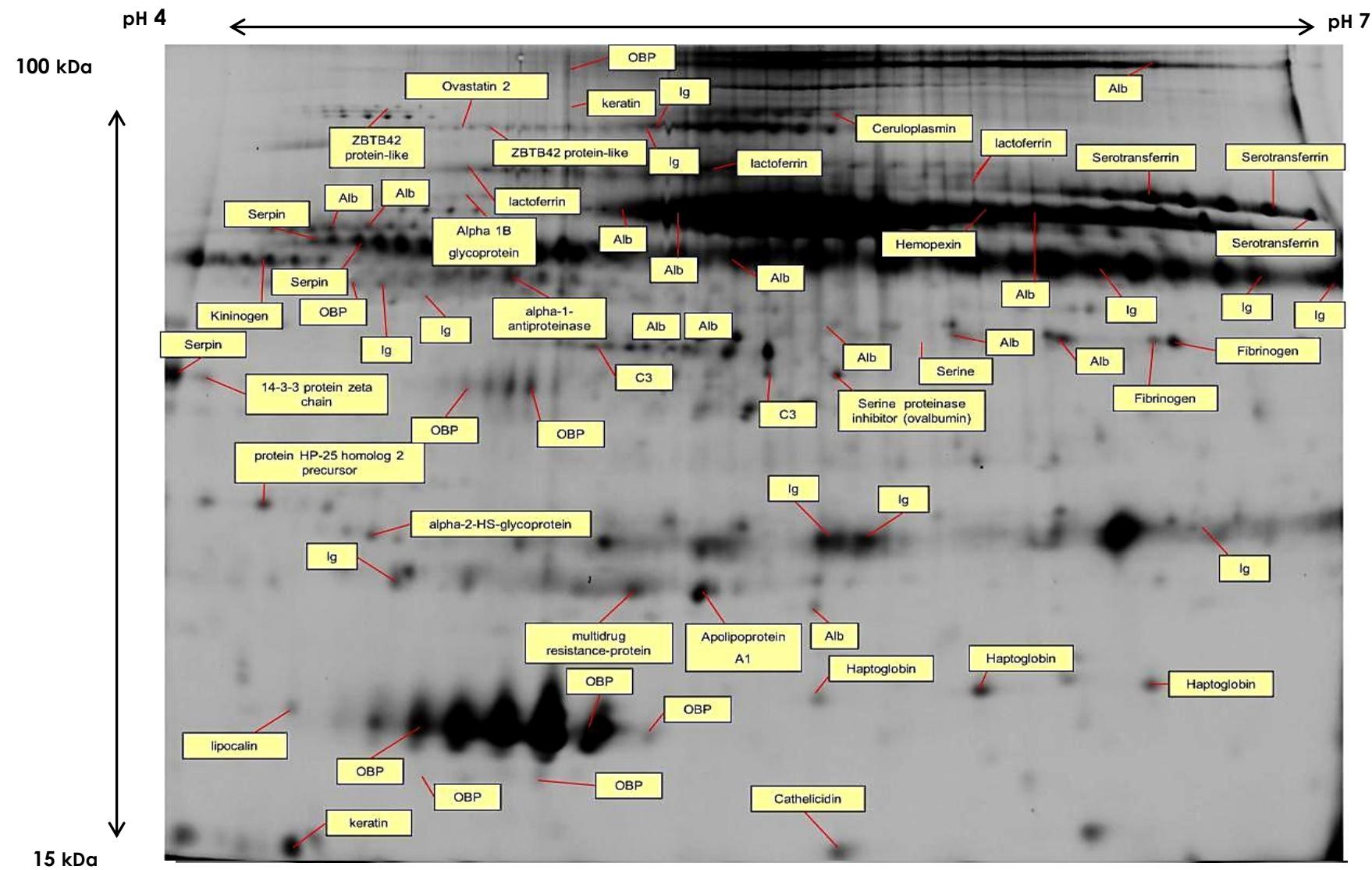
Relative Quantification by Difference Gel Electrophoresis (DiGE) of
Nasal secretion from calves with MCF



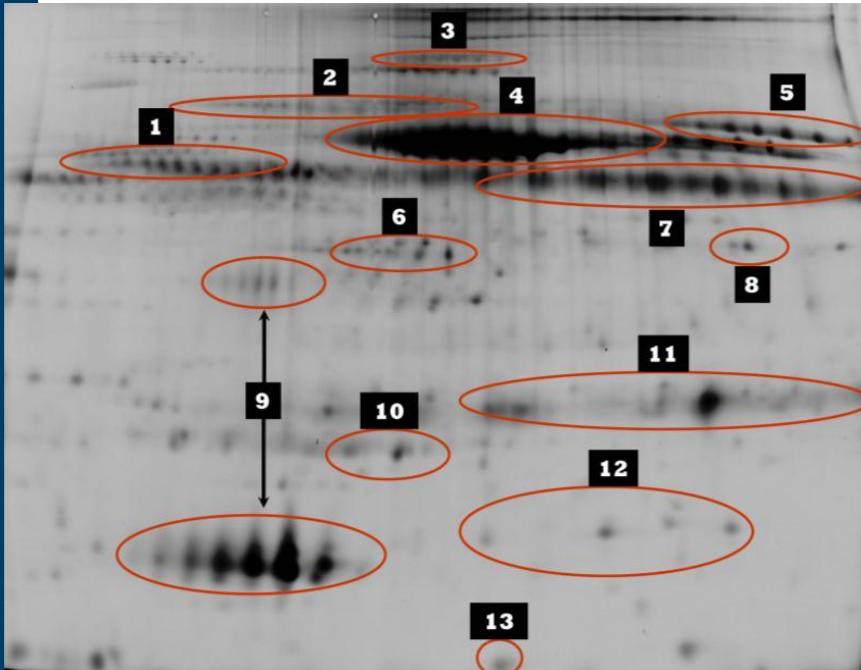




DiGE of Nasal Secretion in MCF

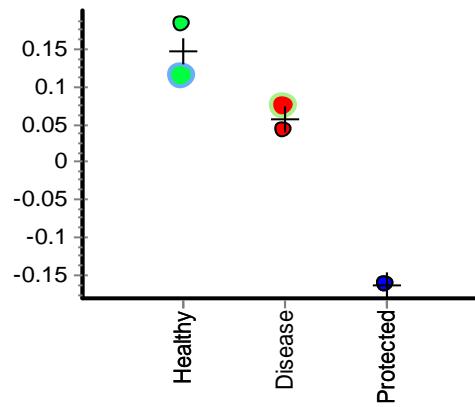


DiGE of Nasal Secretion in MCF

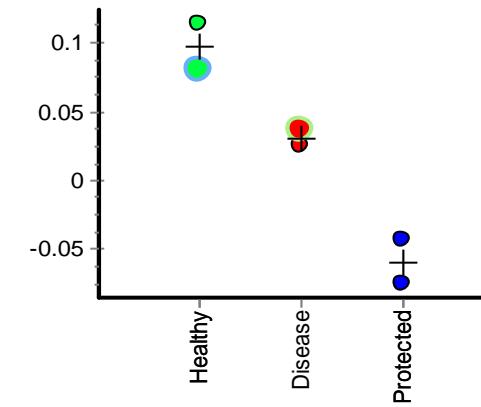


| Protein spot no. | Protein name | Accession | Theoretical mass (Da) | Highest MS score | Highest matches |
|------------------|---------------------------|--------------|-----------------------|------------------|-----------------|
| 1 | Serpin | SPA35_BOVIN | 46482 | 578 | 78 |
| 2 | Lactoferrin | TRFL_BOVIN | 80002 | 1107 | 149 |
| 3 | Ceruloplasmin | NP_001243485 | 121901 | 434 | 40 |
| 4 | Albumin | ALBU_BOVIN | 71244 | 1681 | 228 |
| 5 | Serotransferrin | TRFE_BOVIN | 79870 | 1404 | 147 |
| 6 | Complement C3 (fragments) | CO3_BOVIN | 188652 (~ 68000) | 554 | 110 |
| 7 | Ig heavy chain | O46780_BOVIN | 51391 | 749 | 58 |
| 8 | Fibrinogen beta chain | FIBB_BOVIN | 57146 | 722 | 112 |
| 9 | Odorant-binding protein | OBP_BOVIN | 18492 | 579 | 97 |
| 10 | Apolipoprotein A1 | APOA1_BOVIN | 30258 | 368 | 74 |
| 11 | Ig light chain | Q1RMN8_BOVIN | 25032 | 343 | 45 |
| 12 | Haptoglobin | HPT_BOVIN | 45629 | 383 | 60 |
| 13 | Cathelicidin | CTHL1_BOVIN | 17931 | 412 | 59 |

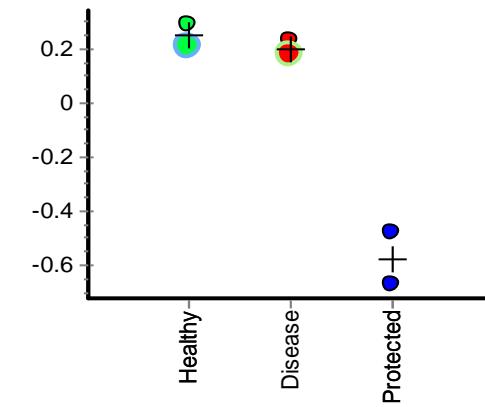
Fibrinogen beta chain



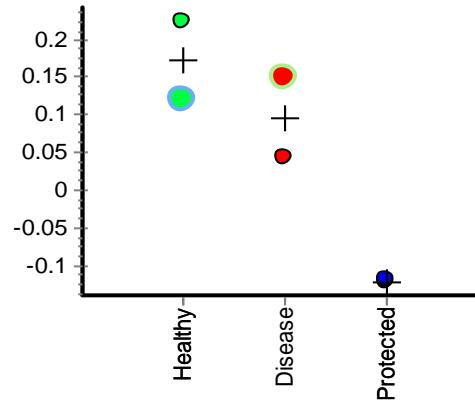
Albumin



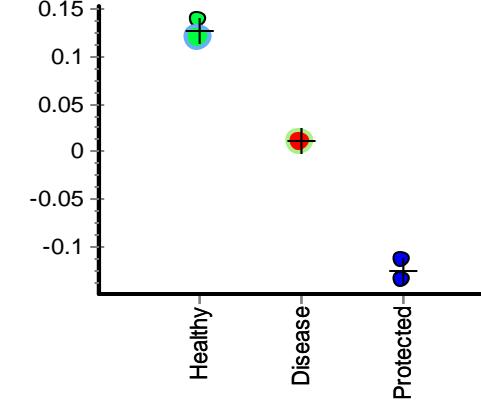
Apolipoprotein A1



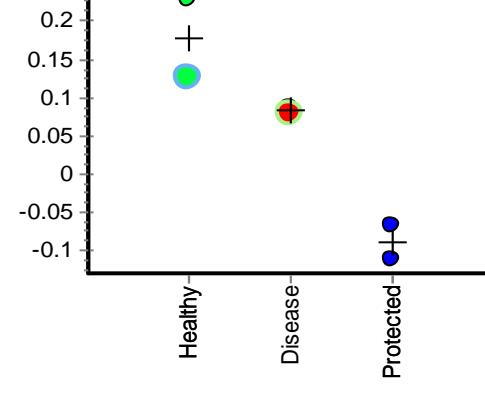
Ceruloplasmin



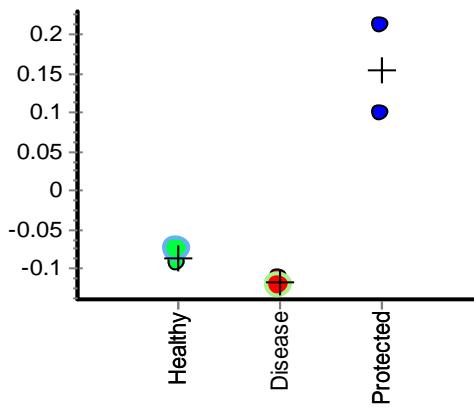
Kininogen



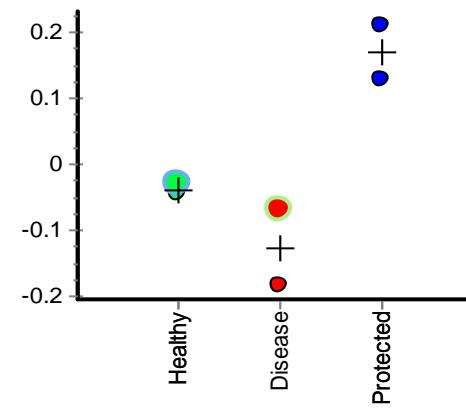
Hemopexin



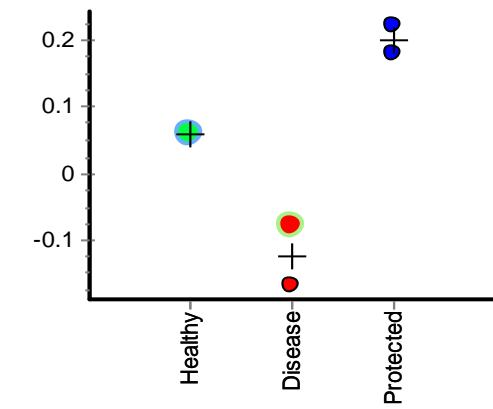
Lactoferrin



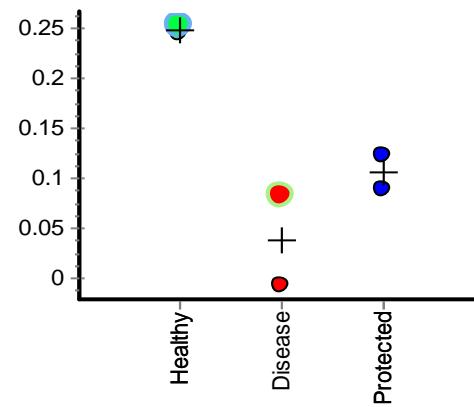
Complement C3



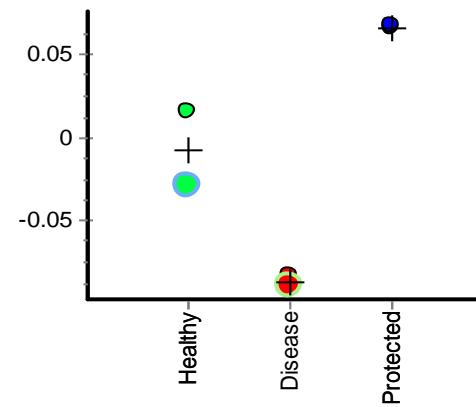
Haptoglobin



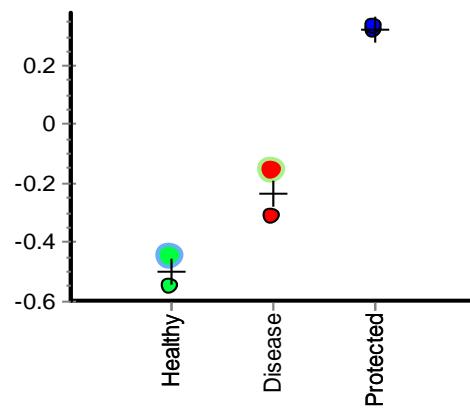
Cathelicidin



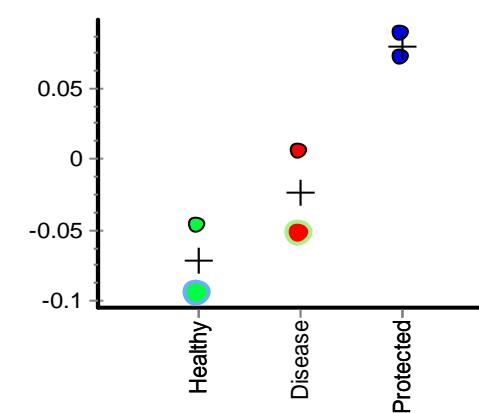
Serotransferrin



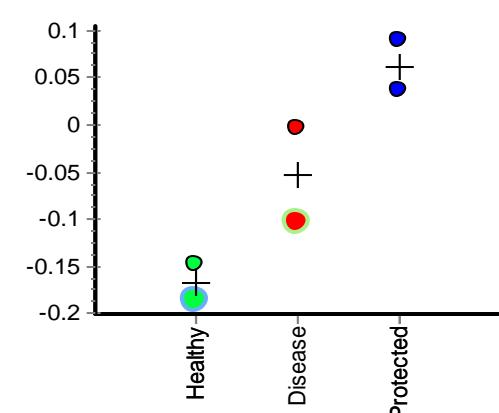
Odorant Binding Protein



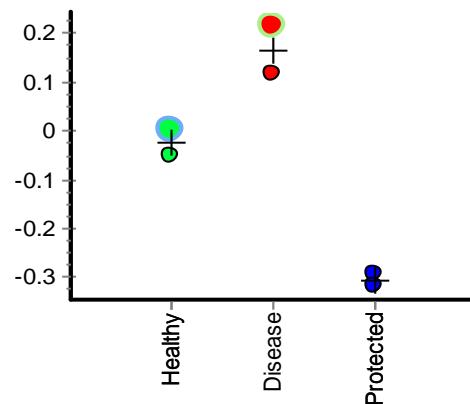
Ig A



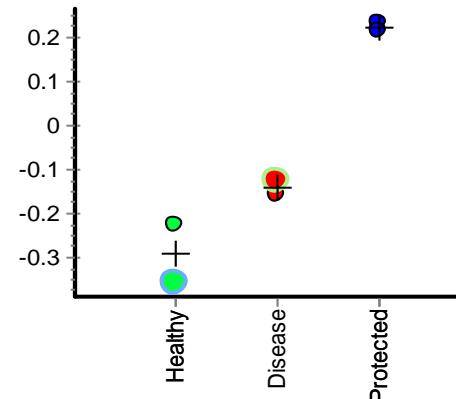
Ig G



Serpin



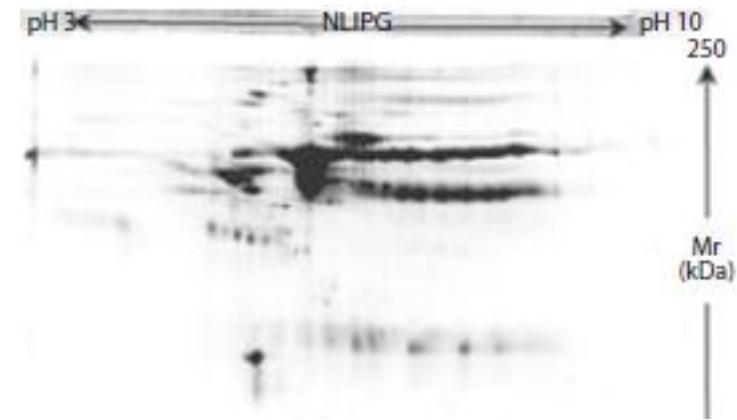
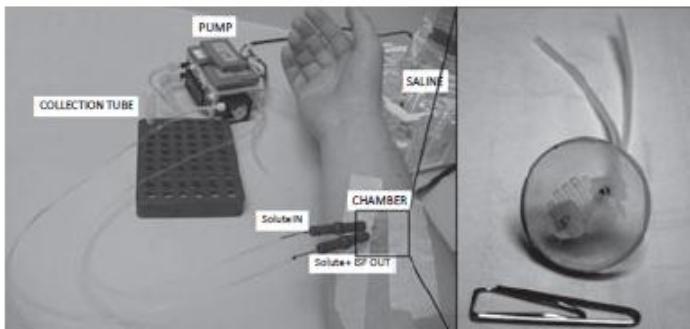
Alpha-2-macroglobulin



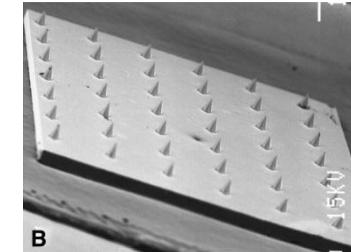
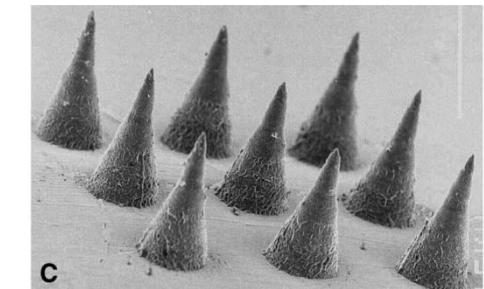
- 23 protein spots were differentially expressed as determined by DiGE
- Most plasma proteins ↓ in challenge cattle
- Immunoglobulins, OBP and Serpin ↑ disease and protected cattle
- Proteins involve in innate immunity and antimicrobial ↑ in protected
↓ in disease

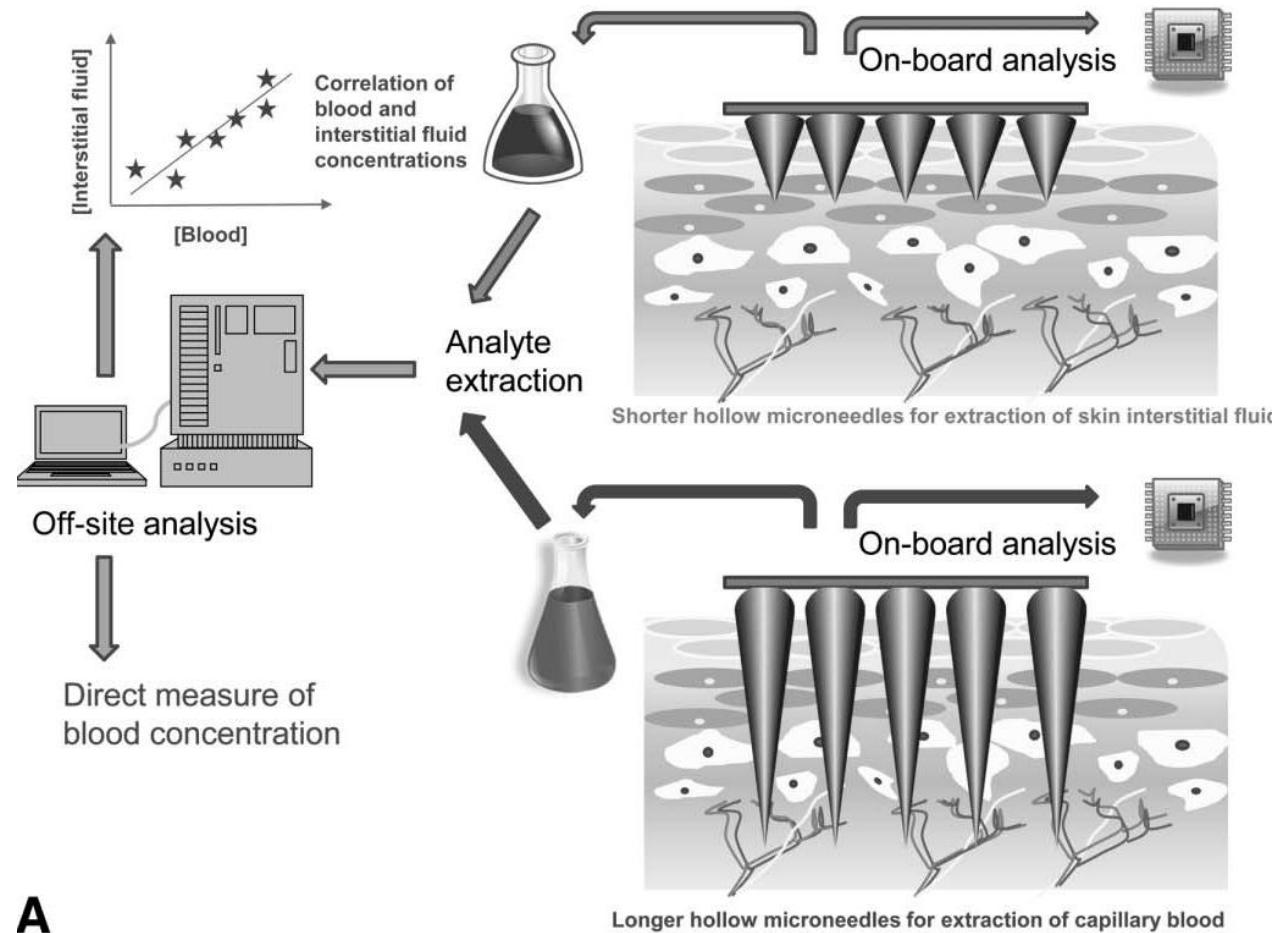
- Device for collecting interstitial fluid 0.5ml of ~1mg/ml protein
- 15min collections up to 90 min
- Protein analysed by 2DE
- Identification underway
- Not suitable for Dairy Care but ISF is a substrate for proteomics

Lecombe et al Skin Res & Tech, 2013 19:27



- Microneedle arrays ~100 per cm²
- Made of polymer (b) poly(methylvinylether-co-maleic acid) and poly(ethyleneglycol) (c) silicone or metal
- For therapeutic drug delivery – also sampling of ISF
- Painless penetration of skin
- Collect interstitial fluid: 50-150 nm
- Collect capillary blood: 1000-1500nm
- Sample collected for analysis
- Potential to link to point of care biosensors



**A**

- Review of microneedle array for glucose monitoring

- ISF collected for continuous glucose monitoring

- Glucose in ISF and serum

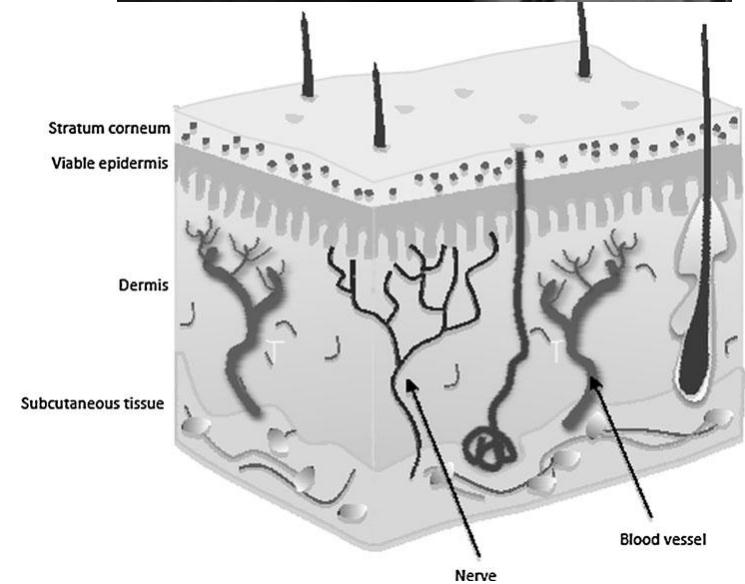
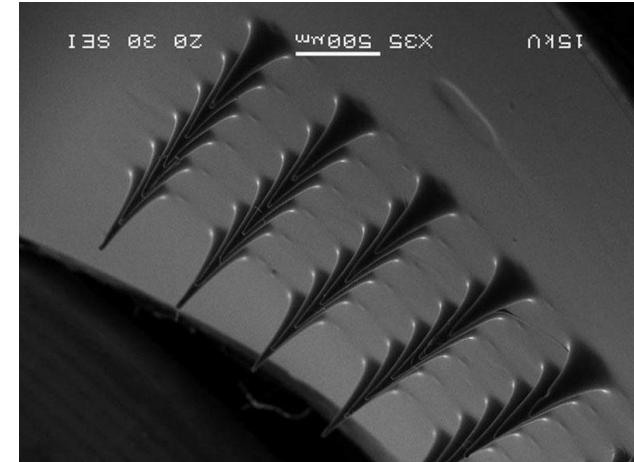
- Pain assessed on visual analog scale

Control 0.42 mm

26 gauge needle 23.9mm

Microneedle array 0.67 mm

- El-Aboudi et al Diabetes Tech Therap 2013
15:101



- **Proteomics can detect changes in the milk protein composition during mastitis**
- **Quantitative proteomics can provide sensitive & multiplexed assays of low abundance milk protein**
- **Peptidomics can provide a panel of peptides which could be used as biomarkers of the cause of mastitis**
- **Metabolomics detects many metabolites in milk, potential as biomarkers of pathogens**
- **Bovine saliva and nasal secretion can be substrate for proteomics**
- **Sweat and interstitial fluid could be collected for omic analysis**

Roshan Mansor
Faizal Ghazali
Emily O'Reilly
Nick Jonsson
Chris Knight
Chris McComb
Rozaihan Mansor
Bill Mullen
Amaya Albalat
David Barrett
Mary Waterston

Support from
BBSRC
EU
Glasgow University
Universiti Putra Malaysia

Mass spectrometry analysis

- **LC system coupled with MS (Dionex UltiMate 1)**
 - separation using a 4.6 x 150mm ZIC-HILIC column at 300µl/min
 - gradient ran from 80% ACN to 20% ACN in 30 min, step to 5% ACN, wash for 9 min, re-equilibrate for 6 min at 80%
- **MS Acquisition (Orbitrap Exactive)**
 - Performed in both –ve and +ve ionisation modes and full scan modes
 - 50 000 resolution, scan range 70- 1400, noise threshold of 5000, m/z tolerance of 0.0005Da, alignment with a mass tolerance of 0.0005 and rt tolerance of 15 sec.